MEDITERRANEAN CORALLIGENOUS ASSEMBLAGES: A SYNTHESIS OF PRESENT KNOWLEDGE

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Abstract   Coralligenous concretions, the unique calcareous formations of biogenic origin in Mediterranean benthic environments, are produced by the accumulation of encrusting algae growing in dim light conditions. This review provides an overview of the results obtained by the main studies dealing with these formations, including the environmental factors which influence the development of coralligenous communities, their distribution, types, assemblages, builders and eroders, the biotic relationships and processes that create and destroy coralligenous assemblages, their dynamics and seasonality, and the functioning of several outstanding and key species. Special attention is devoted to the biodiversity of coralligenous communities and a first estimation of the number of species reported for this habitat is provided. Major disturbances affecting coralligenous communities are discussed, ranging from large-scale events that are probably related to global environmental changes to degradation by waste water or invasive species. Degradation by fishing activities and by divers is also considered. Finally, the main gaps in current scientific knowledge of coralligenous communities are listed and some recommendations are made regarding their protection.

Introduction and description
Encrusting calcareous algae are important components of benthic marine communities within the euphotic zone (Blanc & Molinier 1955, Adey & McIntyre 1973, Littler 1973a, Lebednik 1977, James et al. 1988, Dethier et al. 1991, Adey 1998) and their historical roles as reef builders have been chronicled thoroughly by Wray (1977). Coralline algae are major contributors to coral reef frameworks (Finckh 1904, Hillis-Colinvaux 1986, Littler 1972) where they usually are the dominant reef-forming organisms (Foslie 1907, Odum & Odum 1955, Lee 1967, Littler 1973b). Although encrusting corallines are adapted to grow at low light conditions (Littler et al. 1986, Vadas & Steneck 1988), coralline algal reef frameworks are usually restricted to littoral or shallow sublittoral environments throughout the marine realm (e.g., Littler 1973b, Adey & Vassar 1975, Laborel et al. 1994) because they easily withstand turbulent water motion and abrasion (Littler & Doty 1975, Adey 1978). The only known exception to this restriction is the coralligenous framework, a coralline algal concretion that thrives exclusively in Mediterranean deep waters (20–120 m depth).

There is no real consensus among scientists studying benthic communities in the Mediterranean Sea about what a coralligenous habitat is. In this review a coralligenous habitat is considered to be a hard substratum of biogenic origin that is mainly produced by the accumulation of calcareous encrusting algae growing in dim light conditions. Algae and invertebrates growing in environments with low light levels are called sciaphilic in opposition to photophilic, that is, growing at high light levels. All plants and animals thriving in coralligenous habitats are, thus, sciaphilic. Although more
extensive in the circalittoral zone, coralligenous habitats can also develop in the infralittoral zone, provided that light is dim enough to allow growth of the calcareous algae that produce the calcareous framework. Infralittoral coralligenous concretions always develop on almost vertical walls, in deep channels, or on overhangs, and occupy small surface areas. Communities developing in low light conditions near sea level, in sites of strong water movement and usually below the mediolittoral biogenic rim of the coralline alga *Lithophyllum byssoides* (Boudouresque & Cinelli 1976), are not considered in this review, even though they may exhibit small concretions of coralline algae. Other algal dominated communities thriving in the circalittoral zone, such as rhodolith beds (Basso & Tomaselli 1994) or *Cystoseira zosteroides* assemblages (Ballesteros 1990), are also excluded, as the coralline algal framework in these cases is reduced or almost nil. Some facies of coralligenous communities (and which are categorized as “pre-coralligenous” by several authors, e.g., Pérès & Picard 1964, Gili & Ros 1985, Ros et al. 1985) are also excluded from this review, but only if they refer to sciaphilic communities without a basal framework of coralline algae. Therefore, the main criterion used to define the coralligenous habitat is the presence of a bioherm of coralline algae grown at low irradiance levels and in relatively calm waters. This bioherm is always very complex in structure and, in fact, allows the development of several kinds of communities (Laborel 1961, Laubier 1966), including those dominated by living algae (upper part of the concretions), suspension feeders (lower part of the concretions, wall cavities and overhangs), borers (inside the concretions) and even soft-bottom fauna (in the sediment deposited in cavities and holes). Therefore, the coralligenous habitat should be considered more as a submarine landscape or community puzzle rather than a single community.

**History and main studies**

*Historical account of general and faunal studies*

The word ‘coralligenous’ (*coralligène* in French) was first used by Marion (1883) to describe the hard bottoms that fishermen from Marseilles called *broundo* and which are found at a depth of between 30 and 70 m, below seagrass meadows of *Posidonia oceanica* and above coastal muddy bottoms. *Coralligène* means ‘producer of coral’ and is related to the abundance of red coral (*Corallium rubrum*) found on this type of bottom. Marion (1883) includes long lists of fauna collected in these *coralligène* bottoms. Pruvot (1894, 1895) also used the word *coralligène* to describe similar bottoms in the Pyrenees region of the Mediterranean (Banyuls), and this terminology was included in bionomical descriptions of Mediterranean sea bottoms from the end of the nineteenth century. Feldmann (1937) subsequently described in detail the algal composition of the coralligenous assemblages from Banyuls and identified the main calcareous algae responsible for coralligenous bioherms. He also made observations of the animals contributing to the framework and of bioeroders. Pérès & Picard (1951) continued the work of Marion (1883) on coralligenous bottoms from the Marseilles region, defining the components of the coralligenous assemblages; they demonstrated their high microspatial variability and described the environmental factors which allow them to develop.

Elsewhere in the Mediterranean, Bacci (1947), Tortonese (1958), Rossi (1958, 1961), Parenzan (1960) and Molinier (1960) characterized the pre-coralligenous and coralligenous bioherms in some areas of the Italian coast and Corsica and Pérès & Picard (1958) described the coralligenous communities from the northeastern Mediterranean. The last authors reported several warm-water species, as well as the absence of various species that dominate coralligenous concretions in the western Mediterranean. Laborel (1960, 1961) also expanded the study of coralligenous communities to other Mediterranean areas, including the eastern Mediterranean. He described five main coralligenous types (cave and overhang concretions, wall concretions, concretions at the base of submarine
walls, concretions over flat rocky surfaces and platform coralligenous assemblages) and, in his 1960 paper, also provided the first quantified lists of algal and animal species obtained by scuba diving.

In 1964 Pérès & Picard (1964) summarised existing knowledge of coralligenous communities, defining the notion of pre-coralligenous and simplifying the categories of Laborel (1961) into two coralligenous types: coralligenous assemblages over littoral rock and bank or platform coralligenous assemblages, according to the original substratum (rock or sediment) where concretion began. They proposed an evolutionary series relating the different biocenoses of the circalittoral zone in the Mediterranean and suggested that the coralligenous community was the climax biocenosis of this zone. They also used the word ‘precoralligenous’ to refer to a facies with a great development of erect, noncalcareous, sciaphilic algae and a low cover of invertebrates. An English summary of Pérès & Picard’s (1964) work can be found in Pérès (1967). At about the same time, Vaissière (1964), Fredj (1964) and Carpine (1964) made interesting contributions to the distribution and bionomic description of coralligenous concretions in the region of Nice and Monaco, east of Marseilles.

Gamulin-Brida (1965) conducted the first bionomical studies of coralligenous communities in the Adriatic Sea and concluded that they are biogeographically very similar to those found in the northwestern Mediterranean, with a great abundance of large bryozoans, gorgonians and alcyonarians.

Laubier (1966) made a major contribution to knowledge of invertebrates living in coralligenous assemblages, with his study based on data from the Pyrenean region of the Mediterranean. He was the first to report the high biodiversity of these substrata, he carefully studied the fauna of the concretions (particularly accurate are the studies on polychaetes, copepods and echinoderms) and defined the physico-chemical conditions allowing the coralligenous communities to develop. He was also the first to make a large number of observations related to the natural history of the species inhabiting coralligenous assemblages and, in particular, referred to the relationships of epibiosis, endobiosis, commensalism and parasitism. Subsequent to Laubier’s studies, Sarà (1968, 1969) described the coralligenous communities in the Pouilles region (Italy) and True (1970) collected quantitative samples from the coralligenous assemblages of Marseilles, providing data on the biomass of the main species of suspension feeders.

Hong (1980, 1982) exhaustively described the coralligenous communities from Marseilles and the effects of sewage on their fauna. He also described the animals that contribute to these coralligenous frameworks and defined four different categories of invertebrates which can be distinguished by considering their ecological significance in the assemblages. Extensive lists of several taxonomic groups (mainly foraminiferans, sponges, molluscs, pycnogonids, amphipods and bryozoans) greatly increased the knowledge of the biodiversity of coralligenous communities.

Gili & Ros (1984) reviewed the coralligenous communities of the Medes Islands, off the northeast coast of Spain, and accurately evaluated the total surface area occupied by coralligenous assemblages in this marine reserve (Gili & Ros 1985). Detailed species lists of most algal and animal groups for coralligenous communities from specific areas of the Spanish Mediterranean can also be found in Ballesteros et al. (1993) and Ballesteros & Tomas (1999). Sartoretto (1996) studied the growth rate of coralligenous buildups by radiocarbon dating and related the growth periods to different environmental conditions, mainly the eustatic water level and the transparency of the water column. He also identified the main calcareous algae that finally produce the framework and emphasised the importance of *Mesophyllum alternans*. The effect of sedimentation and erosion by browsers and borers was also quantified.

**Algal studies**

Feldmann (1937) was the first to describe unequivocally the algal composition of coralligenous assemblages; he differentiated these substrata from the deep-water algal beds of *Cystoseira spinosa*.
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and C. zosteroides, and identified the main calcareous algae responsible for coralligenous deposition. The algal community growing on coralligenous assemblages was named the *Pseudolithophyllum expansum-Lithophyllum hauckii* association.

Scuba diving was first used in the study of algal flora of coralligenous assemblages by Giaccone (1965), who made some species lists of coralligenous communities and described a particular plant association, the *Pseudolithophyllum-Halimedetum platydiscae* in the area of Palermo (Sicily). Giaccone & De Leo (1966) also used scuba diving to study the coralligenous and precoralligenous communities of the Gulf of Palermo by using the phytosociological method of Braun Blanquet. They distinguished both types of communities and referred to them as an association of *Lithophyllum expansum* and *Lithothamnion philippi* (coralligenous) and an association of *Halimeda platydisca* and *Udotea desfontainii* (precoralligenous). The population of *Laminaria rodriguezii* growing over a coralligenous community at the island of Ustica was also studied by Giaccone (1967), although this endemic Mediterranean kelp is usually more abundant in deep-water rhodolith beds (*fonds à pralinés*) (Molinier, 1956).

Boudouresque (1970) studied the macroalgal communities of coralligenous concretions as part of a detailed and exhaustive study of the sciaphilic benthic communities in the western Mediterranean. The accurate methodology (Boudouresque, 1971) included scuba sampling and further sorting and identification in the laboratory. Augier et al. (1971) used the same methods to study the algal sciaphilic communities around the island of Port-Cros (France).

Boudouresque (1973) proposed that the terms coralligenous and precoralligenous be avoided, as they have a physiognomical value but do not refer to any bionomical or phytosociological entity; instead, he joined all the sciaphilic algal settlements under relatively sheltered conditions into one association (*Peyssonnelietum rubrae*), and created two subassociations, corresponding to the assemblages developing in the infralittoral zone (*Peyssonnelietum aglaothamnietosum*) and the circalittoral zone (*Peyssonnelietum rodriguezelletosum*). He reported the high biodiversity of these assemblages and defined the ecological group of algae characteristic of coralligenous concretions (CC or *Rodriguezellikon*).

Augier & Boudouresque (1975) argued that the algal composition of coralligenous communities thriving in deep water differs from that of sciaphilic assemblages from the infralittoral zone, and named it *Rodriguezelletum strangulorii* according to phytosociological nomenclature.

Boudouresque (1980) and Coppejans & Hermy (1985) made significant contributions to the study of algal assemblages of coralligenous communities in Corsica, but Ballesteros (1991a,b,c, 1992) was the first to provide data on the dynamics and small-scale structure of algal assemblages from coralligenous communities.

Giaccone et al. (1994) conducted a phytosociological review of sciaphilic assemblages described for the Mediterranean. According to this review, most phytobenthic coralligenous assemblages should be included in the order *Lithophylletalia*, where two associations are distinguished: the *Lithophyllum-Halimedetum tunae* described by Giaccone (1965) and the *Rodriguezelletum strangulorii* described by Augier & Boudouresque (1975). Phytobenthic assemblages growing in coralligenous concretions on vertical walls and overhangs in the infralittoral zone should be included in the order *Rhodymenietalia*, and mainly belong to the association *Udoteo-Peyssonnelietum squamariae* described by Molinier (1960) in Corsica, and which seems to be identical to the association of *Peyssonnelia squamaria* described by Feldmann (1937) for the Pyrenees region of the Mediterranean.

Contributions by Ferdeghini et al. (2000) and Acunto et al. (2001), using photographic sampling, demonstrated the small-scale variability in algal assemblages from coralligenous communities, mainly due to the patchy distribution of calcareous algae and other dominant organisms. Recently, Piazzi et al. (2004) carefully studied the algal composition of coralligenous banks developing in three different subtidal habitats (islands, continental shores and offshore banks), and reported high spatial variability at reduced scales but no major differences between assemblages at a habitat level.
Environmental factors and distribution

Light

Light is probably the most important environmental factor with respect to the distribution of benthic organisms along the rocky bottoms of the continental shelf (Ballesteros 1992, Martí et al. 2004, 2005). It is also very important for the development and growth of coralligenous frameworks, as its main builders are macroalgae which need enough light to grow but which cannot withstand high levels of irradiance (Pérès & Picard 1964, Laubier 1966).

According to Ballesteros (1992), coralligenous communities are able to develop at irradiances ranging from 1.3 MJ m⁻² yr⁻¹ to 50–100 MJ m⁻² yr⁻¹, that is, between 0.05% and 3% of the surface irradiance. Similar ranges are reported by Ballesteros & Zabala (1993), who consider the lower light limit for the growth of Mediterranean corallines to be at around 0.05% of the surface irradiance (Figure 1). These values agree with those obtained by Laubier (1966) in the coralligenous communities of Banyuls, where he reported, at a depth of 32 m, light levels of 1.8–2.6% of surface irradiance at noon in September. However, light levels reaching different microenvironments of coralligenous communities can differ by at least two orders of magnitude. For example, Laubier (1966) reported light levels in an overhang dominated by red coral to be 17-fold lower than those recorded in an exposed, horizontal surface. Light levels reaching small holes and cavities of coralligenous banks must be almost zero, and similar to light levels reaching the bathyal zone or the innermost part of caves.

The quality of light reaching coralligenous bottoms should also be taken into account. Most of the light belongs to the blue and green wavelengths, with green light dominating in relatively murky waters in winter and in coastal continental waters, and blue light dominating in summer and in offshore banks and islands (Ballesteros 1992) (Figure 2). Although most authors consider that light quantity is much more important than light quality in determining algal growth and primary production (e.g., Lüning 1981, Dring 1981), the absolute dominance of red algae in coralligenous assemblages close to their deepest distribution limit points to the ability of phycobilines to capture light in the ‘green window’ (Ballesteros 1992).

![Figure 1](image-url)
Dissolved nutrients in sea water at coralligenous depths follow the annual pattern described for coastal Mediterranean waters, with the highest values in winter and the lowest in summer. The mean annual water nitrate concentration near the coralligenous concretions at depths of 18 and 40 m at Tossa (northwestern Mediterranean) is around 0.6 μmol l⁻¹, with peaks of 1.5 μmol l⁻¹ in winter and undetectable levels in summer (Ballesteros 1992) (Figure 3). Similar values are reported for a station in Cabrera, at a depth of 50 m (Ballesteros & Zabala 1993). However, these values are much lower than those reported from stations situated close to river mouths, such as the coralligenous communities around the Medes Islands, where mean annual values are close to 1 μmol l⁻¹ (Garrabou 1997). Phosphate concentrations are much lower and are always below 0.1 μmol l⁻¹ at

**Figure 2** Distribution by wavelength (uv: ultraviolet, v: violet, b: blue, g: green, y: yellow, r: red) of submarine irradiances relative to surface irradiance for several depths in August (A) and November (B) in waters off Tossa de Mar (northwestern Mediterranean). (From Ballesteros 1992.)
Tossa and Cabrera (mean concentrations around 0.04 μmol l⁻¹ or lower) (Ballesteros 1992, Ballesteros & Zabala 1993), and always below 0.2 μmol l⁻¹ around the Medes Islands (mean concentrations around 0.13 μmol l⁻¹) (Garrabou 1997) (Figure 3). Coralligenous communities seem to be adapted to these low nutrient concentrations in sea water, as increased nutrient availability greatly affects the specific composition, inhibits coralligenous construction, and increases destruction rates (Hong 1980).

Mean annual particulate organic carbon (POC) rates of 387 μg C l⁻¹ are reported for the near-bottom planktonic community at a depth of 15 m around the Medes Islands (Ribes et al. 1999a), although winter and spring values were much higher (500–800 μg C l⁻¹). Dissolved organic carbon (DOC) rates, also reported by Ribes et al. (1999a) for the same site, amount to 2560 μg C l⁻¹, peaking in spring and summer (Figure 4). Ribes et al. (1999a) concluded that the detrital fraction was the dominant component of total organic carbon in the near-bottom planktonic community throughout the year, which could be explained by the importance of runoff particles in the Medes Islands, but may also be due to the input of organic matter by macroalgal (and seagrass) production and the activity of benthic suspension feeders in removing microbial organisms from the plankton. However, further studies are necessary in this regard because the Medes Islands are strongly affected by continental inputs of DOC and POC, which is not usually the case for most Mediterranean coastal areas (mainly in islands or in the southern part).

**Water movement**

Although flowing currents predominate at depths where coralligenous communities develop (Riedl, 1966), water movement generated by waves is very significant even at depths of 50 m (Ballesteros & Zabala, 1993; Garrabou, 1997) for wave heights >1 m. The year-round average of water motion for a coralligenous community in the Medes Islands at a depth of 25–35 m is 40 mg CaSO₄ h⁻¹.
that is, one order of magnitude lower than water motion at a depth of 2 m (Garrabou, 1997) (Figure 5). However, due to the intricate morphology of coralligenous frameworks, water movement can differ greatly between various microenvironments, in a similar way to that reported for light levels (Laubier, 1966).

**Temperature**

Most of the organisms living in coralligenous communities are able to support the normal seasonal temperature range characteristic of Mediterranean waters. Although Pérès & Picard (1951) stated that coralligenous communities display a relative stenothermy, Laubier (1966) described an annual temperature range of 10–23˚C in the coralligenous assemblages of Banyuls. Pascual & Flos (1984)

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**Figure 4** Monthly averages expressed as μg C l⁻¹ of live and detrital carbon (A), live carbon (B) and dissolved organic carbon (C) in waters close to coralligenous concretions around the Medes Islands (northwestern Mediterranean). (From Ribes et al. 1999a. With permission from Oxford University Press.)
found temperatures ranging between 12 and 20°C at the shallowest limit of the coralligenous communities of the Medes Islands (20 m depth), although temperatures ranged from 12–16°C close to their deepest limit (60 m depth) (Figure 6). Ballesteros (1992) reported more or less the same temperatures for the coralligenous assemblages developing at depths of 20 and 40 m at Tossa de Mar between the end of November and the end of June (13–16°C), but differences of up to 9°C in summer, when the thermocline is situated at a depth of around 35 m; however, peak temperatures of 22°C were detected at the end of August at a depth of 40 m. In the Balearic Islands, where coralligenous communities are restricted to waters >40 m deep, water temperature ranges from 14.5–17°C for most of the year, although occasional peaks of 22°C are detected at the end of October, when the thermocline is at its deepest (Ballesteros & Zabala 1993). However, some organisms living in coralligenous assemblages from deep waters seem to be highly stenothermal, as they are never found in shallow waters. This is the case, for example, of the kelp *Laminaria rodriguezii*, which seems to be mainly restricted to depths >70 m and is seldom found between 50 and 70 m, except for in seamounts or upwelling systems (Ballesteros, unpublished data). Moreover, recent (1999) large-scale mortality events of benthic suspension feeders thriving in coralligenous communities have been attributed to unusually long-lasting periods of high temperatures during summer (Perez et al. 2000; Romano et al. 2000), although the ultimate cause of these mortalities remains unclear (possible causes include high temperatures, low food availability, pathogens and physiological stress).
Salinity

The relatively shallow and coastal coralligenous communities of Banyuls and the Medes Islands experience salinity ranges between 37 and 38 (Laubier 1966, Pascual & Flos 1984), although salinity variations for coralligenous assemblages from insular areas should be lower.

Geographical distribution

Coralligenous buildups are common all around the Mediterranean coasts, with the possible exception of those of Lebanon and Israel (Laborel, 1987). According to Laborel (1961), the best developed formations are those found in the Aegean Sea, although the most widely studied banks are those of the northwestern Mediterranean; therefore, most of the data presented here come from this area.

Depth distribution

The minimal depth for the formation of coralligenous frameworks depends on the amount of irradiance reaching the sea bottom. On vertical slopes in the area around Marseilles this minimal depth reaches 20 m, but it is much lower in other zones like the Gulf of Fos, where coralligenous communities are able to grow in shallower waters (12 m) because of the high turbidity of the water related to the Rhône mouth. This minimal depth is displaced to deeper waters in insular areas like Corsica or the Balearic Islands, where water transparency is very high (Ballesteros & Zabala 1993). However, coralligenous frameworks can appear in very shallow waters if light conditions are dim enough to allow a significant development of coralline algae (Laborel 1987, Sartoretto 1994) and they may even occur in the clearest waters like those around Cabrera, where they can be found at a depth of only 10 m in a cave entrance (Martí et al. 2004).

The depth distribution of coralligenous assemblages in subhorizontal to horizontal bottoms for different Mediterranean areas is summarised in Table 1.

Figure 6 Average seawater temperatures for a depth of between 0 and 80 m off the Medes Islands (July 1973–December 1977). Shaded area corresponds to depth of coralligenous outcrops. (From Pascual & Flos, 1984. With permission.)
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Table 1  Depth intervals for the distribution of coralligenous outcrops in different Mediterranean areas

<table>
<thead>
<tr>
<th>Region</th>
<th>Depth (m)</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Banyuls</td>
<td>20–40</td>
<td>Feldmann 1937, Laubier 1966</td>
</tr>
<tr>
<td>Marseilles</td>
<td>20–50</td>
<td>Laborel 1961, Hong 1980</td>
</tr>
<tr>
<td>Medes Islands</td>
<td>20–55</td>
<td>Gili &amp; Ros 1984</td>
</tr>
<tr>
<td>Tossa de Mar</td>
<td>20–60</td>
<td>Ballesteros 1992</td>
</tr>
<tr>
<td>Naples</td>
<td>45–70</td>
<td>Bacci 1947</td>
</tr>
<tr>
<td>Cabrera</td>
<td>50–100</td>
<td>Ballesteros et al. 1993</td>
</tr>
<tr>
<td>Corsica</td>
<td>60–80</td>
<td>Laborel 1961</td>
</tr>
<tr>
<td>Northeastern Mediterranean</td>
<td>70–90</td>
<td>Laborel 1961</td>
</tr>
<tr>
<td>Aegean Islands</td>
<td>90–110</td>
<td>Laborel 1961</td>
</tr>
<tr>
<td>Siculo-Tunisian area</td>
<td>90–120</td>
<td>Laborel 1961</td>
</tr>
<tr>
<td>Southeastern Mediterranean</td>
<td>100–120</td>
<td>Laborel 1961</td>
</tr>
</tbody>
</table>

Structure

Coralligenous types: structure and habitats


Banks are flat frameworks with a variable thickness that ranges from 0.5 to several (3–4) m. They are mainly built over more or less horizontal substrata, and have a very cavernous structure (numerous holes, Laborel 1987) that often leads to a very typical morphology (it has been compared to Gruyère cheese) (Figure 7A). These banks are sometimes surrounded by sedimentary substrata, and Pérès & Picard (1952) argued that they developed from the coalescence of rhodoliths or maërl (coralligène de plateau). However, it is highly probable that these frameworks have almost always grown upon rocky outcrops (Got & Laubier 1968, Laborel 1987) (Figure 7B).

Rims develop in the outer part of marine caves and on vertical cliffs, usually in shallower waters than banks. The thickness of rims is also variable and ranges from 20–25 cm to >2 m; thickness increases from shallow to deep waters (Laborel 1987) (Figure 7C).

In shallow water the main algal builder is Mesophyllum alternans, which builds flat or slightly rounded banks or rims with a foliaceous structure. As the water deepens, other corallines (Lithophyllum frondosum, L. cabiochae, Neogoniolithon mamillosum) become important builders. Shallow water banks are generally covered with populations of green algae Halimeda tuna and Flabellia petiolata (Lithophylo-Halimedetum tunae), which can be so dense that they hide the calcareous algae. However, at greater depths the density of these erect algae decreases and corallines dominate the community (Rodriguezcelletum strafforelli).

Holes and cavities within the coralligenous structure always sustain a complex community dominated by suspension feeders (sponges, hydrozoans, anthozoans, bryozoans, serpulids, molluscs, tunicates) (Figure 7D). The smallest crevices and interstices of the coralligenous buildup have an extraordinarily rich and diverse vagile endofauna of polychaetes and crustaceans, while many attached or unattached animals cover the main macroalgae and macrofauna, swarm everywhere, from the surface to the cavities or inside the main organisms, and thrive in the small patches of sediment retained by the framework.
According to Hong (1982) four different categories of invertebrates can be distinguished with respect to their position and ecological significance in the coralligenous structure:

1. Fauna contributing to buildup, which help develop and consolidate the framework created by the calcareous algae. Several bryozoans, polychaetes (serpulids), corals and sponges constitute this category. They include 24% of the total species number.
2. Cryptofauna colonising the small holes and crevices of the coralligenous structure. They represent around 7% of the species, including different molluscs, crustaceans and polychaetes.
3. Epifauna (living over the concretions) and endofauna (living inside the sediments retained by the buildup), which represent a great number of species (nearly 67%).
4. Eroding species, accounting for only around 1%.

*Algal builders*

Coralline algae are the main coralligenous builders (Laborel 1961, Laubier 1966, Sartoretto 1996). The taxonomy of this group of algae is very difficult to determine and the nomenclature of the
species is constantly changing. Due to their great importance in the construction of coralligenous frameworks several issues regarding the taxonomic status and current nomenclature of the main species are considered here.

The main algal building species, according to Sartoretto (1996) and several other authors (e.g., Feldmann 1937, Péレス & Picard 1964, Boudouresque 1970, Hong 1980, Ballesteros 1991b), has repeatedly been identified as *Mesophyllum lichenoides* (Ellis) Lemoine. However, Cabioch & Mendoza (1998) reported the most common species of the genus *Mesophyllum* growing in coralligenous assemblages to be a different species and named it *Mesophyllum alternans* (Foslie) Cabioch & Mendoza (Figure 8A). Although present in the Mediterranean Sea, *M. lichenoides* does not seem to contribute to coralligenous buildup (Cabioch & Mendoza 1998). Therefore, it is likely that some or most of the reports of *M. lichenoides* as a coralligenous builder actually refer to *M. alternans* (Cabioch & Mendoza, 1998) (Figure 8A).

*Pseudolithophyllum expansum* (sensu Lemoine) has been identified by most authors as being the second most common coralline alga in coralligenous concretions. However, Boudouresque & Verlaque (1978) identified another species, similar to *P. expansum*, and described it as *P. cabiochae*. Later, studies by Woelkerling (1983), Athanasiadis (1987), Woerkerling et al. (1993) and Furnari

**Figure 8** (See also Colour Figure 8 in the insert.) Main red algal building species in coralligenous frameworks. (A) *Mesophyllum alternans*; (B) *Lithophyllum frondosum*; (C) *Lithophyllum cabiochae*; (D) *Neogoniolithon mamillatum*; (E) *Peyssonnelia rosa-marina*. (Photos by the author.)
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et al. (1996) shed some light (but also added further confusion) regarding the name to be applied to the alga called *P. expansum* and/or *P. cabiochae* by Mediterranean phycologists and marine biologists. The last review by Athanasiadis (1999a) suggested that *Pseudolithophyllum* should not be regarded as a different genus to *Lithophyllum* and that the two species growing in coralligenous communities should be named *Lithophyllum stictaeforme* (Areschoug) Hauck [= *Lithophyllum frondosum* (Dufour) Furnari, Cormaci & Alongi; = *Pseudolithophyllum expansum* (Philippi) Lemoine; = *Lithophyllum expansum* sensu Lemoine] (Figure 8B) and *Lithophyllum cabiochae* (Boudouresque & Verlaque) Athanasiadis (Figure 8C). However, according to Marc Verlaque (personal communication), *L. stictaeforme* and *L. frondosum* are not synonyms and the species usually reported as *Pseudolithophyllum expansum* by Mediterranean phycologists should be named *Lithophyllum frondosum*.

Moreover, Woelkerling (1983) recognised the lectotype of *Lithophyllum expansum* Philippi (non Lemoine) as a *Mesophyllum* and considered it to be a heterotypic synonym of *M. lichenoides*. However, a recent study by Cabioch & Mendoza (2003) showed that the lectotype of *Lithophyllum expansum* Philippi is specifically different from *Mesophyllum lichenoides*, *M. alternans* and other Mediterranean species of this genus. They named it *Mesophyllum expansum* (Philippi) Cabioch and Mendoza and it corresponds to the taxa usually identified as *Mesophyllum lichenoides* var. *agariciformis* (Pallas) Harvey by Mediterranean phycologists. As a result of all this confusion it is not possible to determine the extent to which *M. expansum* contributes to coralligenous buildup, although it is likely to make a significant contribution, at least in some places. Another species, *Mesophyllum macroblastum* (Foslie) Adey, has been reported for the coralligenous frameworks in Corsica (Cabioch & Mendoza 2003), and a fifth species (*Mesophyllum macedonis* Athanasiadis) (Athanasiadis 1999b) may also be present in the coralligenous frameworks of the Aegean Sea. According to Marc Verlaque (personal communication), three species of the genus *Mesophyllum* coexist in the coralligenous communities off Marseille (*M. alternans*, *M. expansum*, *M. macroblastum*), suggesting a much greater biodiversity of coralligenous coralline algae than expected.

The alga identified by Feldmann (1937) as *Lithophyllum hauckii* (Rothpletz) Lemoine, a very common coralline in the coralligenous buildups of the Banyuls region, should be named *Neogoniolithon mamillosum* (Hauck) Setchell & Mason (Hamel & Lemoine 1953, Bressan & Babbini-Benussi 1996) [= *Spongites mamillosa* (Hauck) Ballesteros] (Figure 8D). Although not a coralline alga, it should also be pointed out that authors prior to 1975 identified the calcareous *Peyssonnelia* growing in coralligenous communities as being *Peyssonnelia polymorpha* (Zanardini) Schmitz. Boudouresque & Denizot (1975) described a similar species, *Peyssonnelia rosa-marina* (Figure 8E), that is more common than *P. polymorpha* and which also contributes to coralligenous frameworks. Therefore, reports of *P. polymorpha* prior to the description of *P. rosa-marina* should probably be regarded as referring to this latter species or to both entities.

Feldmann (1937) identified the four main calcareous algae responsible for the coralligenous frameworks in the region of Banyuls: *Lithophyllum frondosum* (as *Pseudolithophyllum expansum*), *Neogoniolithon mamillosum* (as *Lithophyllum hauckii*), *Mesophyllum alternans* (as *M. lichenoides*) and *Peyssonnelia rosa-marina* f. *saxicola* (as *P. polymorpha*). The same species have also been reported for coralligenous frameworks studied in several areas close to the Gulf of Lions (e.g., Boudouresque 1973, Ballesteros 1992). It seems that these species are almost always the same, with the possible exception of *Lithophyllum frondosum* which seems to be replaced by *L. cabiochae* in several areas of the Mediterranean that are warmer than the Gulf of Lions (e.g., Corsica, Balearic Islands, the eastern Mediterranean).

Hong (1980) reports three species as being the main coralligenous builders in the region of Marseilles: *Lithophyllum cabiochae*, *Mesophyllum alternans* (?) and *Neogoniolithon mamillosum*. *Peyssonnelia rosa-marina* is also very abundant. Other calcareous species contributing to buildup are *Archaeolithothamnion mediterraneum*, *Lithothamnion sonderi* (?) and *Peyssonnelia polymorpha*.
According to Sartoretto et al. (1996), *Mesophyllum alternans* (as *M. lichenoides*) is the main algal building species for both ancient and recent coralligenous constructions in the northwestern Mediterranean. *Mesophyllum alternans* is a highly tolerant species in terms of light, temperature and hydrodynamism, and is currently the dominant species in shallow waters. In some areas, *Peyssonnelia rosa-marina* and *P. polymorpha* may also be the dominant species, and form a very cavernous, highly bioeroded coralligenous framework. In deep waters *Lithophyllum cabiochae* is the main calcareous alga in the region of Marseilles and Corsica, but its cover can vary from one geographical area to another. For example, the encrusting algal cover in deep-water coralligenous frameworks in Marseilles is limited to a few isolated small living thalli that seem insufficient to allow current renewal of the coralligenous construction. In contrast, these deep frameworks are luxuriant in Corsica, as evidenced by the accumulation of living thalli of *L. cabiochae*.

The identification of the species present in the algal framework of coralligenous blocks from 7700 years ago to the present has shown that no species changes have occurred (Sartoretto et al. 1996). The study by Sartoretto et al. (1996) in the Marseilles region and Corsica identified five Corallinaceae and one Peyssonneliaceae: the nongeniculate corallines *Mesophyllum alternans* (as *M. lichenoides*), *Lithophyllum* sp. (as *Titanoderma* sp., probably *Lithophyllum pustulatum v. confinis*), *Lithophyllum cabiochae-frondosum* (discrimination between *L. cabiochae* and *L. frondosum* is uncertain in fossil material), *Lithothamnion* sp., the geniculate coralline alga *Amphiroa verruculosa*, and, finally, *Peyssonnelia* sp. *Mesophyllum alternans* is also the main algal builder in the coralligenous frameworks of the Mediterranean Pyrenees (Bosence, 1985), along with *Lithophyllum* and *Titanoderma* (quoted as *Pseudolithophyllum* and *Tenarea* in Bosence’s paper). *Peyssonnelia polymorpha* and *P. rosa-marina f. sasicola* may also be abundant in the coralligenous frameworks of the Mediterranean Pyrenees, the northeast coast of Spain, and the Balearic Islands (Bosence 1985, Ballesteros 1992, Ballesteros et al. 1993). However, even if *Peyssonnelia* is abundant as a living encrusting alga, it is almost completely absent from the fossil record (Bosence 1985, Sartoretto 1996). Carbonate content of the *Peyssonnelia* species is lower than the average carbonate content in corallines (Laubier 1966, Ballesteros 1992), and calcification in the form of aragonite rather than calcite prevents a good fossilization of these species (James et al. 1988). However, these and other species of *Peyssonnelia* usually have a basal layer of aragonite that may contribute to the consolidation of coralligenous frameworks when mixed with the physico-chemical precipitations of CaCO₃ (Sartoretto 1996).

**Animal builders**

Coralligenous animal builders have been studied in the Marseilles region (Hong 1980) where 124 species contribute to the frameworks, and account for around 19% of the total number of species reported. The most abundant animal group are the bryozoans, accounting for 62% of species, followed by the serpulid polychaetes with 23.4%. Minor contributors are the cnidarians (4%), molluscs (4%), sponges (4%), crustaceans (1.6%) and foraminiferans (0.8%). However, Laborel (1987) considers the foraminiferan *Miniacina miniacea* (Figure 9A) to be the most important animal builder. Hong (1980) distinguished three different types of animal builders: those contributing directly to the framework, and which are relatively large; those with a reduced builder activity due to their small size; and those which agglomerate carbonate particles. The first group includes the bryozoans *Schizomavella* spp., *Onychocella marioni*, *Cribularia radiata*, *Pentapora fascialis*, *Enthalophorovacia deflexa*, *Celleporina caminata*, *Myriapora truncata*, *Brodiella armata* and *Turbi-cellepora coronopus* (Figures 9B,C), several serpulids (*Serpula vermicularis*, *S. concharum*, *Spirobranchus polyrema*) (Figure 9D), the molluscs *Vermetus* sp., *Serpulorbis arenarius* and *Clavagella melitensis*, and the scleractinians *Hoplangia durotrix*, *Leptopsammia pravoti*, *Caryophyllia inornata* and *C. smithii* (Figure 9E). Among the second group, Hong (1980) reports some small bryozoans...
such as *Crassimarginatella maderensis* and *Mollia patellaria*, serpulids like *Hydroides* spp., *Filograna* spp., and *Spirorbis* spp., the cirripedes *Verruca Stróma* and *Balanus perforatus*, and the foraminiferan *Miniacina miniacea*. In terms of the ‘agglomerative’ animals, he reports sponges such as *Geodia* spp., *Spongia virgulosa* and *Fuchsospongia cavernosa*, the bryozoans *Beania* spp., and the alcyonarian *Epizoanthus arenaceus*.

**Bioeroders**

Feldmann (1937) described the abundance of several organisms that erode calcareous concretions, in particular the excavating sponge *Cliona viridis* (Figure 10A), the bivalve *Lithophaga lithophaga* and several annelids. Hong (1980) listed 11 bioeroders in the coralligenous communities of Marseilles: four species of sponges of the genus *Cliona*, three species of molluscs, two species of polychaetes of the genus *Polydora* and two sipunculids. According to Sartoretto (1996), the organisms that erode coralligenous frameworks are similar to those eroding other marine bioherms such
as the *trottoir* of *Lithophyllum byssoides* or the coral reefs. Three types of eroding organisms can be distinguished: browsers, microborers and macroborers.

The only browsers in the coralligenous concretions are sea urchins (Laubier 1966), because the only important Mediterranean fish grazing on algae (*Sarpa salpa*) do not usually thrive in coralligenous communities. *Sphaerechinus granularis* (Figure 10B,D) is an important biological agent that substantially erodes coralligenous concretions, although local variations in sea urchin abundance and individual size greatly influence the amount of calcium carbonate eroded annually. Another sea urchin commonly found in coralligenous communities is *Echinus melo* (Figure 10C). The proportion of calcareous algae in its digestive content ranges from 18–50% of the total (Sartoretto 1996) and it preys mainly on sponges, bryozoans and serpulid polychaetes. Given the low densities of this sea urchin in coralligenous communities (1–3 individuals in 25 m²), Sartoretto (1996) concludes that the bioerosional role of *E. melo* is very limited.

Microborers include blue-green algae (cyanobacteria), green algae and fungi (Hong 1980). Three green algae (*Ostreobium queketti, Phaeophila sp.* and *Eugomontea sp.* ) and four cyanobacteria (*Plectonema tenebrans, Mastigocoleus testarum, Hyella caespitosa* and *Calothrix sp.*), together with some unidentified fungi, seem to be the main microborers in coralligenous communities. Diversity is higher in shallow waters, whereas, according to colonisation studies conducted by Sartoretto (1998), it is restricted to only one species (*Ostreobium*) in deep waters (>60 m).

Macroborers comprise molluscs (*Lithophaga lithophaga, Gastrochaena dubia, Petricola lithophaga, Hyatella arctica*), sipunculids (*Aspidosiphon mülleri, Phascolosoma granulatum*), polychaetes (*Dipolydora spp., Dodecaceria concharum*) and several excavating sponges (Sartoretto 1996, Martin & Britayev 1998). Among perforating sponges commonly found in coralligenous communities, some of them excavate mainly in *Corallium rubrum* and other calcareous cnidarians (*Aka labyrinthica, Scantilletta evispira, Dotona pulchella spp. mediterranea, Cliona janitrix*), whereas others, such as *Pione vastifica, Cliona celata, C. ampicavata, C. schmidti* and *C. viridis* can be...
found in a wide range of calcareous substrata (coralline algae, bivalves, madreporids, etc.) (Rosell & Uriz 2002). Cliona viridis is the most powerful destructive sponge of calcareous substrata (Rosell et al. 1999), and is the most abundant excavating sponge in coralligenous communities (Uriz et al. 1992a). The encrusting sponges and the Sipunculida become more abundant in polluted coralligenous environments (Hong 1983).

Assemblages

The final result of the builders and eroders of coralligenous concretions is a very complex structure, in which several microhabitats can be distinguished (Figure 11). Environmental factors (e.g., light, water movement and sedimentation rates) can vary by one to two orders of magnitude in parts of the same concretion situated as close as one metre from each other. This great environmental heterogeneity allows several different assemblages to coexist in a reduced space. For practical purposes those situated in open waters (from horizontal to almost vertical surfaces) are distinguished here from those situated in overhangs and cavities. The assemblages of macroborers are not discussed because the only available data have already been commented on, nor are the assemblages thriving in the patches of sediment between or inside coralligenous frameworks because there are no quantitative data on them.

Algae, both encrusting corallines and green algae, usually dominate in horizontal to subhorizontal surfaces (Figure 12), although their abundance decreases with depth or in dim light. Phycologists have distinguished two main communities according to the light levels reaching coralligenous frameworks. In shallower waters Mesophyllum alternans usually dominates in the basal layer and Halimeda tuna in the upper stratum, with an important coverage of other algae (Peyssonnelia spp., Flabellia petiolata) (Figure 13A). This plant association has received the name of Lithophyllum-Halimedetum tunae, and has been described in detail by Ballesteros (1991b). Algal biomass ranges between 1200 and 2100 g dry weight (dw) m⁻², while percent cover ranges from 180–400%. The number of species is very high (average of 76 species in 1024 cm²) and average diversity is 2.5 bits ind⁻¹. Its bathymetric distribution ranges from a depth of 12–15 m to 30–35 m in the Gulf of Lions, but it can reach depths below 50 m in the clear waters of seamounts and insular territories of the western and eastern Mediterranean. This association develops at irradiances ranging from around

Figure 11 (See also Colour Figure 11 in the insert.) Diagrammatic section of a coralligenous bank, showing the high small-scale environmental heterogeneity and the different microhabitats. (Drawing by J. Corbera.)
2.3–0.3 W m$^{-2}$, which correspond, respectively, to 3 and 0.4% of the surface irradiance. Other quantified species lists are described in Marino et al. (1998).

In deeper waters or lower irradiances the density of *Halimeda tuna* decreases and other calcareous algae become dominant (*Lithophyllum frondosum, Neogoniolithon mamillosum, Peyssonnelia rosa-marina*) (Figure 14). Other common algae are members of the family Delesseriaceae

Figure 12 (A) Drawing of a coralligenous concretion dominated by algae in the Medes Islands (NE Spain).
and other laminar red algae (*Kallymenia, Fauchea, Sebdenia, Rhodophyllis, Predaea*), as well as the encrusting green alga *Palmophyllum crassum*. These assemblages correspond to the Rodriguezelletum strafforellii of Augier & Boudouresque (1975), which may be identical to the algal assemblage described by Feldmann (1937) for coralligenous concretions from the Mediterranean Pyrenees (Figures 13B,C,D). Quantified species lists can be found in Boudouresque (1973), Augier & Boudouresque (1975), Ballesteros (1992) and Marino et al. (1998). Algal biomass averages 1600 g m⁻² and percent cover 122%, mostly corresponding to encrusting algae and, around 90%, corresponding to corallines; the number of species is low (38 species in 1600 cm² or lower) (Ballesteros 1992).

Animal assemblages of these two plant associations can differ greatly from one to the other, as well as between sites and geographical areas. The abundance of suspension feeders mainly depends on average current intensity and availability of food (plankton, POC, DOC). In the richest zones (e.g., Gulf of Lions, Marseilles area) gorgonians can dominate the community (Figure 15A,B), but in very oligotrophic waters (e.g., Balearic Islands, eastern Mediterranean), sponges, bryozoans and small hexacorals are the dominant suspension feeders (Figure 15C). The only available quantified biomass data of invertebrate assemblages are those of True (1970) gathered from the Marseilles area, and those results are summarized below.

True (1970) studied an assemblage dominated by *Eunicella cavolinii*. He reports a basal layer of encrusting algae accompanied by erect algae (total biomass of 163 g dw m⁻²). *E. cavolinii* is the most abundant species (up to 304 g dw m⁻²), followed by the bryozoans *Pentapora fascialis* (280.1 g dw m⁻²), *Turbiticellepora avicularis* (49.1 g dw m⁻²), *Celleporina caminata* (22.3 g dw m⁻²) and *Myriapora truncata* (19.9 g dw m⁻²). Other less abundant species include unidentified Serpulidae, anthozoans *Parerythropodium coralloides, Alcyonium acaule, Leptopsammia pruvoti* and

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**Figure 13** (See also Colour Figure 13 in the insert.) Different assemblages of algal-dominated coralligenous banks and rims; (A) with *Halimeda tuna* and *Mesophyllum alternans* (Tossa de Mar, NE Spain, 28 m depth); (B) with *Lithophyllum frondosum* (Tossa de Mar, NE Spain, 40 m depth); (C) with *Peyssonnelia rosa-marina, Mesophyllum alternans, Palmophyllum crassum* and *Peyssonnelia squamaria* (Scandola, Corsica, 50 m depth); (D) detail of C. (Photos by the author.)
Figure 14 (See also Colour Figure 14 in the insert.) (A) Drawing of a deep-water, animal-dominated, coralligenous assemblage in the Medes Islands (NE Spain).
Figure 14 (continued) (See also Colour Figure 14 in the insert.) (B) Key to major species, left from top to bottom: *Paramuricea clavata*\(^6\), (and on it *Halecium halecinum*\(^{15}\), *Pteria hirundo*\(^{22}\)), *Agaophenia septifera*\(^{14}\), *Cliona viridis*\(^3\), *Ancyronia acuta*\(^5\), *Acanthella acuta*\(^4\), *Lithophyllum frondosum*\(^1\), *Agelas ovoides*\(^6\), *Palinus elephas*\(^{24}\), *Parazoanthus axinellae*\(^{15}\), *Spirastrella cunctatrix*\(^6\), *Chondrosia reniformis*\(^6\), *Petrosia ficiformis*\(^4\) (and on it *Smittina cervicornis*\(^{23}\) and *Discodoris atracmaulata*\(^{25}\)), *Serpula vermicularis*\(^{21}\), *Caryophyllia inornata*\(^{20}\), *Halocynthia papillosa*\(^{24}\), *Clathrina coriacea*\(^5\), *Corallium rubrum*\(^{14}\) and *Chromis chromis*\(^{32}\) Right, from top to bottom (excluding the above-mentioned species): *Anthias anthias*\(^{31}\), *Eunicella singularis*\(^{15}\), *Diplodus sargus*\(^{29}\), *Codium bursa*\(^{8}\), *Epinephelus marginatus*\(^{30}\), *Phyllangia mouchezii*\(^{24}\), *Galathea strigosa*\(^{25}\), *Synthecium evansi*\(^{13}\), *Dysidea avara*\(^{10}\). (Drawing by M. Zabala & J. Corbera.)
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Figure 15 (See also Colour Figure 15 in the insert.) Different assemblages of animal-dominated coralligenous banks and rims: (A) with gorgonians *Paramuricea clavata* and *Eunicella cavolinii* but also green algae *Halimeda tuna* and *Flabellia petiolaris* (Gargalo, Corsica, 45 m depth); (B) with *Paramuricea clavata* and encrusting sponges in deep waters (Cabrera, Balearic Islands, 65 m depth); (C) with sponges, bryozoans and anthozoans (Cabrera, Balearic Islands, 50 m depth); (D) overhangs with *Corallium rubrum* (Palazzu, Corsica, 35 m depth). (Photos by the author.)

*Caryophyllia smithii*, tunicates *Microcosmus polymorphus* and *Halocynthia papillosa*, foraminiferan *Miniacia miniacea*, sponges *Chondrosia reniformis* and *Axinella damicornis* and other bryozoans (*Adeonella calveti*, *Beania hirtissima*, *Sertella* spp., *Schizomavella* spp. and *Cellaria salicornioides*). The number of collected invertebrate species amounted to 146 in 7500 cm², with a total weight of invertebrates close to 1563 g dw m⁻². The main biomass corresponded to the phylum Bryozoa, closely followed by Cnidaria, and, with much lower values, Annelida, Porifera, Chordata (tunicates) and Mollusca.

Another assemblage studied by True (1970) is that dominated by *Paramuricea clavata*. Populations of *P. clavata* are abundant in steep rocky walls, but they also grow in horizontal to subhorizontal surfaces if light levels are very low. The basal layer of the community can be mainly occupied by algae (usually attributable to *Rodriguezellatum strafforellii* association) or by other suspension feeders (sponges and bryozoans). The lists of True (1970) do not report any algae. *Paramuricea clavata* has a total biomass of 746 g dw m⁻², followed by the cnidarians *Caryophyllia smithii* (326.3 g dw m⁻²) and *Hoplagia durotrix* (188.1 g dw m⁻²), the bryozoan *Celleporina caminata* (119.6 g dw m⁻²), the anthozoan *Leptopsammia pruvoti* (54.9 g dw m⁻²), the bryozoans *Adeonella calveri* (32.8 g dw m⁻²) and *Turbicellepora avicularis* (31.4 g dw m⁻²), and red coral (*Corallium rubrum*, 16.9 g dw m⁻²). Other less abundant species include unidentified Serpulididae, sponges *Ircinia variabilis* (fasciculata in True, 1970), *Spongia officinalis*, *Sarcotragus spinosula*, *Cacospongia scalaris*, *Petrosia ficoiflora*, *Aplysina cavernicola*, *Eurylus euastrum* and *Agelas oroides*, the bryozoan *Sertella septentrionalis*, the alycianarian *Parazoanthus axinellae*, molluscs *Pteria hirundo*, *Serpulorbis arenarius*, *Lithophaga lithophaga* and *Anomia ephippium*, and tunicates *Microcosmus polymorphus* and *Polycarpa pomaria*. The number of collected invertebrate species
amounts to 111 in 7500 cm², with a total weight of 3175 g dw m⁻². The main biomass corresponds to the phylum Cnidaria, followed by Annelida, Bryozoa, Porifera, Mollusca and Chordata.

Gili & Ballesteros (1991) described the species composition and abundance of the cnidarian populations in coralligenous concretions around the Medes Islands that are dominated by the gorgonian *Paramuricea clavata*. Total cnidarian biomass amounted to 430 g dw m⁻², with 13 species of hydrozoans and 9 species of anthozoans found in an area of 5202 cm². Species contributing the most to the total biomass of the taxocoenosis were the anthozoans *Paramuricea clavata*, *Leptopsammia pruvoti*, *Parazoanthus axinellae*, *Caryophyllia inornata*, *C. smithii*, *Alcyonium acaule* and *Parerythropodium coralloides*, the hydrozoans *Sertularella gaudichaudii* and *Halecium tenellum* also being abundant.

Overhangs and big cavities of coralligenous assemblages have a different species composition to that found in open waters (Figure 15D). Algae are usually completely absent because light is very reduced. However, some thalli of encrusting corallines, *Peyssonnelia* spp. and *Palmophyllum crassum*, can occasionally be found. There are no quantified species lists for this kind of habitat reported in the literature except for those of True (1970), which, in fact, do not come from a coralligenous buildup but from a semidark zone dominated by red coral in a cave (Grotte de l’Île Plane). This assemblage is worth describing as it is very similar to those that develop in the overhangs of coralligenous constructions in the northwestern Mediterranean, or in coralligenous communities situated in very deep waters.

The assemblage of red coral described by True (1970) is dominated by the cnidarians *Corallium rubrum* (2002 g dw m⁻²), *Caryophyllia smithii* (303 g dw m⁻²), *Hoplangia durotrix* (54.1 g dw m⁻²) and *Leptopsammia pruvoti* (52.4 g dw m⁻²), the sponges *Petrosia ficiformis* (241.5 g dw m⁻²) and *Aplysina cavernicola* (27.9 g dw m⁻²), the bryozoan *Celleporina caminata* (100.5 g dw m⁻²), and unidentified Serpulidae (232.4 g dw m⁻²). Other abundant species are the sponges *Ircinia variabilis*, *Spongia officinalis*, *Aaptos aaptos* and *Ircinia oros*, the molluscs *Chama gryphoides* and *Anomia ephippium*, and several unidentified bryozoans. The total number of identified invertebrate species is 63 in 7500 cm², with a total biomass of 3817 g dw m⁻². The dominant phylum is largely the Cnidaria, although Porifera, Annelida and Bryozoa are also abundant.

It should be remembered that most of the invertebrate data presented in this chapter, if representative at all, reflect the biomass and species composition of several assemblages of coralligenous buildups from the Gulf of Lions, which are different to those reported from other sites of the western Mediterranean (e.g., Balearic Islands; Ballesteros et al. 1993) or the eastern Mediterranean (Péres & Picard 1958, Laborel 1960). Therefore, these data cannot be extrapolated to the whole Mediterranean.

**Biodiversity**

Coralligenous communities constitute the second most important ‘hot spot’ of species diversity in the Mediterranean, after the *Posidonia oceanica* meadows (Boudouresque 2004a). However, there appear to be no previous estimates of the number of species that thrive in these coralligenous assemblages. Furthermore, due to their rich fauna (Laubier 1966), complex structure (Péres & Picard 1964, Ros et al. 1985), and the paucity of studies dealing with coralligenous biodiversity, they probably harbour more species than any other Mediterranean community. In fact, coralligenous assemblages are one of the preferred diving spots for tourists due to the great diversity of organisms (Harmelin 1993). Divers are astonished by the high number of species belonging to taxonomic groups as diverse as sponges, gorgonians, molluscs, bryozoans, tunicates, crustaceans or fishes. Moreover, there are innumerable organisms living in these coralligenous communities that cannot be observed by diving, nor without a careful sorting of samples. For example, in a sample of 370 g dw of *Mesophyllum* from a small coralligenous concretion in the south of Spain, García-Raso...
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(1988) found 903 specimens of crustaceans, molluscs and polychaetes; other organisms from other groups (pycnogonids, nematodes, echinoderms, sipunculids, sponges, tunicates, small fishes, such as Gobiidae and Blenniidae, as well as hydrozoans and bryozoans) were also abundant, although not quantified.

Laubier (1966) was the first author to emphasize the great biodiversity of coralligenous communities and listed 544 invertebrates from coralligenous assemblages in the region of Banyuls. Later, in an exhaustive survey of coralligenous communities around Marseilles, Hong (1980) listed a total of 682 species, while several authors (in Ros et al. 1984) report 497 species of invertebrates in the coralligenous assemblages of the Medes Islands. Estimates of the species richness found in coralligenous communities give a very conservative number of 1241 invertebrates (Table 2). Boudouresque (1973) has estimated that at least 315 species of macroalgae can thrive in Mediterranean sciaphilic communities (the coralligenous type being the most widespread). Finally, there are no estimates of the number of fishes that can be found in coralligenous communities, due to the high mobility of most species of this group, but estimates based on available literature regarding the biology of Mediterranean fishes (e.g., Whitehead et al. 1984–1986, Corbera et al. 1996, Mayol et al. 2000) range between 110 and 125 species.

It is very difficult to mention all the species found to date in coralligenous communities, as the existing taxonomic literature is huge and contains many synonyms; this makes it impossible for a nonspecialist in most of the groups to come up with an accurate number of reported species. Nevertheless, an attempt is made at a first, and very conservative, estimate of the total number of species, which amounts to some 1,666 (Table 2). A first step toward increased knowledge of the biodiversity present in coralligenous communities would be to obtain a more accurate estimate of which species have been found and their number.

The next section describes the main findings reported for each taxonomic group.

Taxonomic groups

Algae

At least 315 species of macroalgae thrive in deep-water Mediterranean sciaphilic communities (Boudouresque 1973), and most of them are found in coralligenous concretions. The algal assemblages found here show high biodiversity, with an average of 40 algal species in 600 cm².

Boudouresque (1973) defined the ecological group of algae characteristic of coralligenous concretions (CC or Rodrigezellikon), which (Boudouresque, 1985) comprises 28 species (e.g., Rodriguezella spp., Aeodes marginata, Fauchea repens, Chondrymenia lobata, Gulsonia nodulosa, Polysiphonia elongata, Neogoniolithon mamillosum). However, coralligenous communities are never dominated by this group of species, but rather by other species with a more depth-related widespread distribution, examples being the encrusting corallines Mesophyllum alternans, Lithophyllum frondosum, and L. cabiochae, the green algae Palmophyllum crassum, Flabellia petiolata, Halimeda tuna and Valonia macrophysa, some brown algae such as Dictyota dichotoma, Dictyopteris polyiodoideae, Spatoglossum solierii, Zonaria tournefortii, Halopteris filicina, Phyllariopsis brevipes, Zanardinia prototypus and Laminaria rodriguezii, and a large number of red algae (several species of Peyssonnelia, Kallymenia, Halymenia, Sebdenia, Predaea, Euapogon, Myriogramme, Neurocaulon foliosum, Acrodiscus vidovichii, Osmundaria volubilis, Phyllophora crispa, Rhodymenia ardissonii, Acrosorium venulosum, Rhodophyllis divaricata, Hypoglossum hypoglossoides, Polysiphonia banyulensis, Plocamium cartilagineum, Sphaerococcus coronopifolius, Erythrogressum sandriatum, and Aglaosthannion tripinnatum) (Boudouresque 1973, 1985, Ballesteros 1992, 1993).

The algal component of coralligenous communities largely consists of Mediterranean endemics, which quantitatively represent between 33 and 48% of the total flora (Boudouresque 1985).
Coralligenous communities are rich in algal species, although this richness is lower than that found in photophilic or moderately sciaphilic communities (Ballesteros 1992). Ballesteros (1991b) reports 90 species of macroalgae from the coralligenous assemblages of Tossa de Mar, where *Mesophyllum alternans* and *Halimeda tuna* dominate, but only 38 in the coralligenous communities from a deep water site (Ballesteros 1992). Piazzi et al. (2004) found small differences between algal assemblages of coralligenous habitats along the coast of Tuscany (Italy). However, algal

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populations in coralligenous habitats differ greatly on geographical scales across the whole Mediterranean (Boudouresque 1973) and this is the main reason why, even if the species diversity at one site is rather constant, the overall algal richness of coralligenous habitats — on a Mediterranean-wide scale and covering all depths where they are present — can be huge.

Protozoa

Fifty-four species of Foraminifera are listed by Hong (1980) in the checklist of species from the coralligenous communities of Marseilles, although none of these species seems to be characteristic of coralligenous habitats. *Miniacina miniacae* is the most abundant species, and other common species include *Massilina secans, Planorbulina mediterraneensis, Elphidium crispum* and *Triloculina rotunda*. Laubier (1966) reports six species of Folliculinaeae living as epibionts of bryozoans.

Porifera

Coralligenous communities are very rich in sponges, which grow mainly in the more sciaphilic environments but also in more exposed areas. There are also some species (Clionidae) that are active bioeroders and which excavate the coralline framework. The number of species reported from different well-studied areas is 26 species from Banyuls (Laubier 1966), 78 species from Marseilles (Hong 1980), 48 species from the Medes Islands (Bibiloni et al. 1984), 74 species from Cabrera (Ballesteros et al. 1993), and 24 species from Tossa (Ballesteros & Tomas 1999). The list of sponges reported in all these studies (along with those of True 1970 and Rosell & Uriz 2002) amounts to 142 different species. According to Hong (1980) the following species are characteristic of coralligenous biocoenoses: *Axinella damicornis, Acanthella acuta, Hymedesmia pansa, Agelas oroides, Dictyonella pelligera, Haliclonia mediterranea, Spongionella pulchella* and *Faciospongia cavernosa*. Other abundant sponges (Laubier 1966, True 1970, Hong 1980, Bibiloni et al. 1984, Ballesteros et al. 1993, Ballesteros & Tomas 1999) are: *Cliona viridis, Clathrina clathrus, Oscarella lobularis, Chondrosia reniformis, Phorbas tenacior, Geodia cyanonium, Aaptos aaptos, Pleraplysilla spinifera, Dysidea avara, Terpios fugax, Spongia virgultosa, S. agaricina, S. officinalis, Ircinia variabilis, I. oros, Axinella verrucosa, A. polypoides, Diplastrella bistellata, Petrosea ficiformis, Hexadella racovitzai, Cacospongia scalaris, Dictyonella obtusa, Erylus euastrom, Hippoposophgia communis, Reniera cratera, R. fulva, R. mucosa, Spirastrella canctatrix, Spongiosorites intricatus and Hemimycale lumbrum*.

The coralligenous communities from the eastern Mediterranean seem to be very rich in sponges (Pérès & Picard 1958) because they are almost devoid of alcyonarians and gorgonians. The most abundant species have already been cited above. Those of the genus *Axinella* (*A. polypoides, A. damicornis, A. verrucosa*), *Agelas oroides* and *Petrosea ficiformis* (Pérès & Picard 1958) are particularly common.

Hydrozoa


The only detailed study of hydrozoans found on coralligenous assemblages is that of Llobet et al. (1991a), who report 35 species of hydroids living on the thalli of *Halimeda tuna* in the coralligenous concretions of Tossa de Mar (northwestern Mediterranean). Llobet et al. (1991a)
classify the most abundant hydrozoans into three categories on the basis of their horizontal zonation on the thalli. The hydroids common on the proximal articles (oldest) are relatively large and present throughout the year (*Eudendrium racemosum*, *E. capillare*, *Halecium tenellum* and *Kirchenpaueria echinulata*). Those common on the medial articles (*Campalecium medusiferum*, *Halecium pusillum*, *Hydranthea margarica*, *Phialella quadrata*, *Campanularia everta* and *Filellum serpens*) are smaller and often occur in dense monospecific patches. Finally, those common on the distal articles (*Campanularia raridentata*, *Clytia hemisphaerica*, *Sertularia distans*, *Sertularella polyzonias* and *Aglaphenia pluma*) are present for only short periods and are highly opportunistic. This zonation seems to reflect interspecific niche selection, enabling successful competition for space with other hydroids, algae and bryozoans.

**Anthozoa**


**Scyphozoa**

The only species reported (Hong, 1980) is *Nausitoë punctata*, living inside several massive sponges.

**Turbellaria**

Laubier (1966) reports three turbellarians from the coralligenous communities of Banyuls, all very rare.

**Nemertea**

Nemerteans live endolithically in concretions. According to Pruvot (1897) and Laubier (1966), who report up to 12 species in the coralligenous communities of Banyuls, they are rather common. *Drepanophorus crassus*, *Tetramastix coronatum*, *Micrura aurantiaca* and *M. fasciolata* are the most abundant.

**Nematodes**

Nematodes are the most abundant microscopic metazoans in marine sediments and are present in the sediments retained in coralligenous assemblages, as well as in the endofauna of concretions and the epifauna of algae and sessile invertebrates. However, there are no studies dealing with this group of organisms in coralligenous assemblages.

**Polychaeta**

Polychaetes are extremely abundant in coralligenous communities. Martin (1987) reported a total of 9195 individuals present in 20 samples of 400 cm² collected from coralligenous communities dominated by *Mesophyllum alternans* and *Lithophyllum frondosum* from the Catalan coast (northwestern Mediterranean). This means an average of 460 worms per sample and a density of more than one individual per cm². He found 191 species, with a dominance of Syllidae (31% of the total). The number of species per sample was very high, ranging between 32 and 71 for macrofauna.
(>0.4 mm) and between 27 and 55 for microfauna (<0.04 mm). Diversity of the samples was also very high, averaging 4.54 bits ind\(^{-1}\) for macrofauna and 4.34 bits ind\(^{-1}\) for microfauna (Shannon-Weaver index). According to Martin (1987), coralligenous assemblages are a very suitable habitat for polychaetes because the high structural complexity of the concretions allows the coexistence of several species in a reduced space.

The first checklist of polychaetes collected from coralligenous communities and studied by a specialist is that of Bellan (1964), who reported 70 species. Laubier (1966) reported 130 species in the polychaete assemblages of two coralligenous stations in the Banyuls region; *Lepidasthenia elegans*, *Kefersteinia cirrata*, *Xenosyllis scabra* and *Typosyllis variegata* were the most abundant. According to his observations, and those of Bellan (1964), the polychaetes inhabiting coralligenous concretions are mainly ubiquitous species, although he distinguished two main groups: microfauna and macrofauna. Microfauna comprise three ecological groups: psammophilic species (e.g., *Xenosyllis scabra*, *Eurysillis tuberculata*, *Trypanosyllis coeliaca*), limic species (e.g., *Scalibregmatidae*, *Sclerocheilus minutus*, *Syllides longocirrata*). Among the macrofauna he distinguished four ecological groups: polychaetes living inside sponges (e.g., *Lepidasthenia elegans*, *Eunice siciliensis*, *Amphitrite variabilis*); species living in small crevices and holes, like most Serpulidae and Terebellidae, as well as *Eunice torquata*; big vagile polychaetes living over or inside coralligenous holes (e.g., *Lepidonotus clava*, *Harmothoe aerolata*, *Pontogenia chrysocoma*, *Trypanosyllis zebra*) and, finally, excavating species of the genus *Dipolydora* and *Dodecaceria concharum*.

Hong (1980) reported a total of 109 species of polychaetes inhabiting the coralligenous communities of Marseilles, and distinguished some characteristic species such as *Haplosyllis spongicola*, *Trypanosyllis coeliaca*, *Platynereis coccinea*, *Eunice torquata*, *Lumbrinereis coccinea* and *Potamilla reniformis*. According to Martin (1987), who studied polychaete fauna in the coralligenous communities from the Catalan coast, the most dominant and constant species are *Filograna implexa*, *Spirobranchus polytrema*, *Polydora caeca*, *Pomatoceros triqueter*, *Nereis pelagica*, *Syllis truncata*, *S. gerlachi*, *Haplosyllis spongicola*, *Serpula concharum*, *Anaitides muscosa* and *Dodecaceria concharum*. However, the most conspicuous species growing in coralligenous communities are not usually the most abundant, but rather the large and very apparent species of serpulids (True 1970) notably *Salmacina dysteri*, *Serpula vermicularis*, *S. concharum*, *Sabella pavonina*, *S. spallanzani*, *Myxicola aesthetica* and *Protula* spp. (Ballesteros & Tomas 1999).

**Sipunculida**

Always endolithic, the most abundant species of sipunculid is *Phascolosoma granulatum*, which, along with *Aspidosiphon mülleri*, is also a very active bioeroder (Sartoretto 1996). Laubier (1966) reports a third species in the coralligenous community of Banyuls: *Golfingia minuta*.

**Echiura**

*Bonnellia viridis*, very common in coralligenous communities, is an important detritus feeder. Laubier (1966) reports another, extremely rare species from the coralligenous community of Banyuls (*Thalassema* sp.).

**Mollusca**

Molluscs are extremely abundant in coralligenous communities. Martin et al. (1990) reported a total of 897 individuals in 20 samples of 400 cm\(^2\), equivalent to an average of 45 species per sample.
and more than one mollusc per 10 cm². They report a very high number of species given the reduced area they sampled: 131. The number of species per sample ranged between 5 and 33. Average diversity for all the samples was 3 bits ind⁻¹ (Shannon-Weaver index). Salas & Hergueta (1986) also reported a very high diversity, with an average of 22.7 species per sample.

The number of species reported in studies devoted to the coralligenous communities of a small geographic area are always high: 69 species in Banyuls (Laubier 1966), 142 species in Marseilles (Hong 1980) and 108 species in the Medes Islands (Huelin & Ros 1984). According to these authors, and to Martin et al. (1990), the most common and constant species are the chiton Callochiton achatinus; the prosobranchs Acmaea virginea, Calliostoma zizyphinum, Alvania lineata, A. cancellata, Setia semistriata, S. tenera, Chauvetia minima, C. mamillata, Hinia incrassata, Fusinus pulchellus, F. rostratus, Raphitoma linearis, Clanculus corallinus, Rissoina bruquierei, Triphora perversa, Muricopsis cristatus and Bittium reticulatum; the opisthobranchs Odostomia rissoides, Diaphorodis papillata, Limacia clavigera, Cadlina laevis, Hyspelodoris fontandrai, Chromodoris luteocea, C. purpurea, Dendrodoris grandiflora, Duvaucelia striata, Discodoris atromaculata, Glossodoris gracilis, G. tricolor, Polycera quadrilineata, Flabellina affinis and Donicide banyulensis and the bivalves Arca barbata, Striaria lactea, Musculus costulatus, Kellia suborbicularis, Lithophaga lithophaga, Coralliphaga lithophagella, Anomia ephippium, Pteria hirundo, Chlamys multistriata, Chama gryphoides, Lima lima and Hiatella arctica.

Cephalopods are also present in coralligenous communities, although they are usually not reported in lists. Both Octopus vulgaris and Sepia officinalis are regularly present. Loligo vulgaris eggs are frequently seen in late winter and early spring in some coralligenous platforms.

Acarif

Mites are always rare in coralligenous communities. Laubier (1966) reports six species from Banyuls.

Pycnogonida

Up to 15 species of pycnogonids occur in the coralligenous communities of Marseilles (Hong 1980). Achelia echinata, Rynchothorax mediterraneus, Tanystylum conirostre and Callipallene spectrum seem to be the most common, although they are always rare. Only one species is reported by Laubier (1966) from Banyuls, and two species by Munilla & De Haro (1984) from the Medes Islands.

Copepoda

The fauna of copepods has been carefully studied by Laubier (1966) in one station from the coralligenous communities of Banyuls. He reports up to 54 species. Ectinostoma dentatum, Harpacticus littoralis, Tisbe furcata, Thalestris rufioleScens, Phyllolesteris mysis, Dactylopodia tisboides, Diosaccus tenuicornis, Amphiascus minutus, A. cinctus and Laophonte cornuta are extremely abundant. There are several copepods which live as parasites of different invertebrates: polychaetes, sponges, echinoderms, molluscs, cnidarians and tunicates (Laubier 1966 and references therein).

Ostracoda

Although several species of ostracods are present in coralligenous communities (Laubier 1966, Hong 1980), no study has been devoted to this group. Laubier (1966) reports more than 10 unidentified species in the ‘endogean’ microfauna.
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Cirripedia

The barnacles *Balanus perforatus* and *Verruca strömia*, in coralligenous walls and crevices, and *Acasta spongites*, an endobiont of several sponges (e.g., *Dysidea, Ircinia*), have been identified in coralligenous communities (Laubier 1966, Hong 1980, Carbonell 1984).

Phyllocarida

Only one species has been recorded in the coralligenous communities of Marseilles (Hong 1980).

Mysidacea

Hong (1980) reports seven mysids from the coralligenous communities of Banyuls.

Cumacea

Three cumaceans are reported from coralligenous communities and are always rare (Laubier 1966, Hong 1980).

Tanaidacea

*Tanais cavolini* and *Leptochelia savignyi* are rather common among the ‘endogean’ microfauna of coralligenous frameworks (Laubier 1966, Hong 1980).

Amphipoda

A noteworthy number of amphipods have been sampled in coralligenous communities. Although Laubier (1966) only reports 12 species from the coralligenous communities of Banyuls, a list of 49 species is given by Hong (1980) in Marseilles, and 40 species are reported by Jimeno & Turon (1995) in an extensive survey of the concretions by *Mesophyllum alternans* along the coast of Catalonia (northwestern Mediterranean).

Coralligenous assemblages harbour a certain number of amphipods from photophilic algal communities, together with rheophobic and sciaphilic species, which are linked to the presence of hydroids, sponges and bryozoans. Bellan-Santini (1998) lists 44 species from the coralligenous community (below 35 m depth), to which another 56 species collected from sciaphilic communities with *Flabellia petiolata* and *Halimeda tuna* have to be added. Therefore, a total number of 100 species is probably a good estimate of the amphipods thriving in coralligenous communities.

According to the available literature, common species include *Maera inaequipes, M. grossimana, Liljeborgia dellavallei, Leptocheirus bispinosus, Gitana sarsi, Amphilochochus picudarus, Colomastix pusilla, Iphimedia serratipes* and *Stenothoe tergestina*. In coralligenous communities with some erect algae, the following species are also abundant: *Orchomene humilis, Leptocheirus guttatus, Stenothoe dolfusi, Leucothoe venetiarum, Pseudoprotella phasma, Cressa cristata, C. mediterranea, Caprella acanthifera, Corophium sextonae, Dexamine thea, Leucothoe euryonyx, Aora spinicornis* and *Elasmopus vachoni*. Few species (Harpinia ala, Tryphosella similime, Uncionella lunata) have been collected solely in coralligenous communities (Bellan-Santini 1998).

Isopoda

Laubier (1966) and Hong (1980) report 14 species from coralligenous communities. *Cymodoce truncata, Jaeropsis brevicornis, Paranthura nigropunctata, Synisoma sp., Gnathia maxillaris* and *Paragnathia formica* seem to be relatively common species.
Decapoda

The density of decapods in coralligenous concretions is very high, the estimate being 170 individuals in 500 g dw of *Mesophyllum alternans* (García-Raso & Fernández Muñoz 1987). According to García-Raso et al. (1996), it is very difficult to distinguish characteristic species of the coralligenous community because the assemblages are very similar to those found in other communities where there is shelter (e.g., the rhizomes of *Posidonia oceanica*).

*Alpheus dentipes, Athanas nitescens, Pilumnus hirtellus, Pisidia longicornis, Galathea bolivari, Cestopagurus timidus* and *Thoralus cranchii* are considered to be the characteristic decapod crustaceans inhabiting the shallow coralligenous frameworks of *Mesophyllum alternans* in the southwestern Mediterranean, along with, in certain areas, *Porcellana platycheles, Synalpheus hululensis* and *Calcinus tubularis* (García-Raso 1988). The three species which account for most of the biomass of the decapod crustaceans in the shallow coralligenous communities of the southwestern Mediterranean use this environment in different ways. In *Pilumnus hirtellus*, the coralligenous habitat seems to be a recruitment site, where mainly juveniles are recorded. The whole life cycle of *Alpheus dentipes* takes place in the coralligenous concretions, whereas in *Synalpheus hululensis* the coralligenous habitat provides shelter only for reproductive individuals (García-Raso & Fernández Muñoz 1987).

Other species of decapods frequently reported from coralligenous bottoms are *Alpheus ruber, A. megacheles, Pilumnus spinifer, Pisa tetraodon, Galathea intermedia, Eurynome aspera, Macropodia cerniavskii, Inachus thoracicus, Processa macrophthalma, Periclimenes scriptus, Typton spongicola, Balsia gasti* and *Pisidia longimana* (Laubier 1966, Hong 1980, Carbonell 1984, García-Raso 1988). Other large decapods that are usually found in coralligenous communities are *Dromia personata, Palinurus elephas, Scyllarus arctus, Scyllarides latus* and *Homarus gammarus* (Corbera et al. 1993).

In deep waters, the decapod fauna reported by García-Raso (1989) is different from that reported from shallow water coralligenous habitats. This author found a total of 30 species, with *Pilumnus inermis, Galathea nexa* and *Euchirograpsus liguricus* being the most abundant decapods in these kinds of bottoms from the southwestern Mediterranean.

Pterobranchia

Only one pterobranch, *Rhabdopleura normani*, is reported by Laubier (1966) living as an epibiont of bryozoans.

Brachiopoda

Brachiopod species usually inhabit small crevices and interstices within the concretionary masses of the coralligenous assemblages. *Crania anomala, Argyrothea cistellata, A. cordata, A. cuneata, Megathiris detruncata* and *Lacazella mediterranea* are the brachiopods most commonly reported from coralligenous communities (Laubier 1966, Logan 1979, Hong 1980). Another two species, *Megerlia truncata* and *Platidia davidsoni*, which are more typical of the bathyal zone, are seldom collected from coralligenous habitats (Vaissière & Fredj 1963, Gamulin-Brida 1967, Logan 1979).

Bryozoa

Bryozoans are very abundant in coralligenous communities: 67 species in Banyuls (Laubier 1966), 133 in Marseilles (Hong 1980), 113 in the Medes Islands (Zabala 1984) and 92 in Cabrera (Ballesteros et al. 1993). A tentative estimate of the total number of bryozoans that thrive in coralligenous bottoms according to these studies is around 170 species.
According to Zabala (1986) four different aspects of the distribution of bryozoans can be distinguished in coralligenous communities. The main species mentioned below derive from the studies by Laubier (1966), Hong (1980), Zabala (1984, 1986) and Ballesteros et al. (1993).

1. The flat surfaces of the coralligenous platform are dominated by *Pentapora fascialis* and *Myriapora truncata*, which have *Nolella* spp., *Aetea* spp., *Crissa* spp., *Scrupocellaria* spp., *Mimosella verticillata* and *Synnotum aegyptiacum* as epibionts. *Turbicellepora avicularis* is very common overgrowing gorgonians, and *Chorizopora brongniartii*, *Diplosolen obelium*, *Tabulipora plumosa*, *Puellina guttata* and *Lichenopora radiata* are common epibionts of other organisms. Other common species are *Beania magellanica*, *B. hirtissima*, *Mollia patellaria*, *Schizomavella auriculata*, *Cellepora pumicosa*, *Plagioecia* spp., *Cellaria fistulosa* and *C. salicornioides*.

2. Coralligenous walls have the species reported above but also *Smittina cervicornis*, *Adelonella calveti*, *Chartella tenella*, *Cribilaria innominata*, *Schizomavella* spp., *Parasmittina tropica*, *Sertella* spp., *Caberea boryi* and *Spiralaria gregaria*.

3. Cavities and overhangs of coralligenous outcrops reveal a bryozoan fauna that is almost identical to that present in semidark caves, with several species already reported above, along with *Dentiporella sardonica*, *Brodella armata*, *Turbicellepora coronopus*, *Rynchozoa bispinosum*, *Schizotheca serratimargo*, *Escharoides coccinea*, *Escharina vulgaris*, *Callopora dumerilii*, *Smittoidea reticulata*, *Cribilaria radiata*, *Hippomenella mucronelliformis*, *Crassimarginatella madrensis*, *C. crassimarginata*, *Buskea nitida*, *Celleporina* spp., *Prenantia inerma*, *Diaporoecia* spp., *Enthalophoroecia deflexa* and *Idmidronea atlantica*.

4. A final group is made up of species that appear mainly in deep-water coralligenous habitats, below 50 m depth, and are composed of stenotherm species that are also very resistant to sedimentation: *Figularia figularis*, *Escharina dutertrei*, *E. porosa*, *Onychocella marioni*, *Omaloseca ramulosa*, *Buskea dichotoma*, *Escharella ventricosa*, *Enthalophoroecia gracilis*, *Schizoporella magnifica*, *Mecynoecia delicatula*, *Idmidronea coerulae* and *Hornera frondiculae*.

**Crinoidea**

Two crinoids have been reported from coralligenous habitats, the common *Antedon mediterranea* (Laubier 1966, Ballesteros et al. 1993) and *A. bifida* (Montserrat 1984).

**Ophiuroidea**

According to Tortonese (1965), Laubier (1966), Hong (1980) and Montserrat (1984), up to 17 species of ophiuroids have been reported from coralligenous communities. There are some species that can be considered as characteristic of these habitats, such as *Ophioconis forbesii*, *Amphiura mediterranea* and *A. apicula* (Tortonese 1965, Laubier 1966). Other brittlestars live entangled in gorgonians: *Astropartus sardonica*, *A. armata*, *Turbicellepora coronopus*, *Rynchozoa bispinosum*, *Schizotheca serratimargo*, *Escharoides coccinea*, *Escharina vulgaris*, *Callopora dumerilii*, *Smittoidea reticulata*, *Cribilaria radiata*, *Hippomenella mucronelliformis*, *Crassimarginatella madrensis*, *C. crassimarginata*, *Buskea nitida*, *Celleporina* spp., *Prenantia inerma*, *Diaporoecia* spp., *Enthalophoroecia deflexa* and *Idmidronea atlantica*.

**Asteroidea**

Up to eight species of seastars have been reported from coralligenous bottoms (Tortonese 1965, Laubier 1966, Munar 1993). The most abundant species is the ubiquitous *Echinaster sepositus*. 

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Marthasterias glacialis and Hacelia attenuata are also common, while Ophidiaster ophidianus is only found in the southern, warmer areas of the Mediterranean.

**Echinoidea**

Fourteen species of sea urchins are reported from coralligenous communities (Tortonese 1965, Laubier 1966, Hong 1980, Montserrat 1984, Munar 1993). The most common species is Sphaerechinus granularis (Sartoretto 1996), which is an important bioeroder. Also common in deep waters are Genocidaris maculata and Echinus melo. Psammechinus microtuberculatus is usually hidden inside the cavities of coralligenous outcrops. Juveniles of Paracentrotus lividus (and Arbacia lixula) are sometimes found, but are never abundant. Centrostephanus longispinus is more abundant in the warmer areas of the Mediterranean and usually lives within coralligenous crevices (Pèrès & Picard 1958, Laboré 1960, Harmelin et al. 1980, Francour 1991). Finally, Echinocycamus pusillus is a ubiquitous and very small species that inhabits the small patches of sand and gravel inside the concretions.

**Holothurioidea**

The most commonly observed species of sea cucumber is Holothuria forskali, which can be rather abundant in some coralligenous platforms (Laubier 1966, Ballesteros & Tomas 1999). However, the genus Cucumaria has several species that live endolithically (C. saxicolus, C. planci, C. kirschenbergii, C. petiti). Another four species typical of sandy and muddy habitats have also been reported (Tortonese 1965, Laubier 1966, Montserrat 1984): Holothuria tubulosa, H. mammata, Trachytyone tergestina and Stichopus regalis.

**Tunicata**

Ramos (1991) describes a high species richness of ascidians in coralligenous communities, the families Didemnidae and Polyclinidae being especially present. In fact, around 70% of ascidian fauna is present in the coralligenous community (82 species). According to Ramos (1991), the most characteristic species of the coralligenous community are Cystodites dellechiajei, Ciona edwardsi and Halocynthia papillosa, although other abundant species include Diplosoma spongiforme, Distaplia rosea, Trididemnum cereum, T. armatum and Polycarpa gracilis. Other species that are often collected from coralligenous communities are Distomus variolosus, Didemnum maculosum, Ecteinascidia herdmanni, Clavelina nana, Polysyncratochiton lacazei, P. bilobatum, Polycarpa pomaria, Pyura spp., Microcosmus polymorphus, M. sabatieri, Styela partita, Eudistoma planum, E. banyulensis, Pseudodistoma cyrnusense, Aplidium densum and A. conicum (Laubier 1966; Hong 1980; Turon 1990, 1993). Clavelina dellavallei and Rhodosoma verecundum seem to be especially abundant in the coralligenous concretions from the eastern Mediterranean (Pèrès & Picard 1958).

**Pisces**

The fish fauna from the coralligenous community includes many fishes covering a wide bathymetric range, such as Epinephelus marginatus, Sciaena umbra, Coris julis, Dentex dentex, Symphodus mediterraneus, S. tinca, Diplodus vulgaris, Apogon imberbis, Chromis chromis or Labrus merula. However, there is a group of species that are characteristic of coralligenous communities. Some of these, like Lappanella fasciata or Acantholabrus palloni, are species restricted to deep waters (Sartoretto et al. 1997), but others, such as Anthias anthias (Harmelin 1990), as well as (among the commonest species) Gobius vittatus, Physic physicus and Labrus bimaculatus (Garcia-Rubies 1993, 1997), are easily observed during recreational diving. Other species are more abundant in
coralligenous outcrops than in shallow waters, examples being *Serranus cabrilla*, *Spondyliosoma cantharus*, *Diplodus punctazzo*, *Ctenolabrus rupestris*, *Spicara smaris*, *Scorpaena scrofa* and *Symphodus doderleini*. Finally, *Conger conger*, *Muraena helena*, *Zeus faber*, *Scorpaena notata*, *Scyliorhinus canicula* and *S. stellaris* are also observed in the coralligenous habitat (Sartoretto et al. 1997, Ballesteros, personal observation).

The fish fauna inhabiting the small crevices of coralligenous concretions probably consists of fishes with cave-dwelling tendencies, although data are very scarce. Hong (1980) reports juveniles of *Diplecogaster bimaculata* and *Gobius niger*. According to Patzner (1999), cryptobenthic species, such as *Thorogobius ephippiatus*, *T. macrolepis*, *Corcyrogobius liechtensteini*, *Gammogobius steinitzii* and *Didogobius splecmnai*, which are usually observed in caves, may also be present in the small holes of deep water coralligenous habitats. *Odondebuenia balearica* is another cryptobenthic fish that inhabits coralligenous communities but is rarely observed (Riera et al. 1993).

Studies of the fish fauna of the coralligenous habitat have obtained slightly different results when performed in different areas (Bell 1983; Harmelin 1990; Garcia-Rubies 1993, 1997; Ballesteros & Tomas 1999). These differences should be related to biogeography or to differences in coralligenous rugosity. *Symphodus melanocercus*, for example, is a characteristic coralligenous species in Cabrera and other localities of the Balearic Islands, but it is a widespread species in terms of depth distribution in the northwestern Mediterranean (Garcia-Rubies 1993).

**Endangered species**

Although it is very difficult to determine the conservation status of any marine species living in the relatively deep waters where coralligenous communities develop, several approaches to endangered species have been taken.

According to Boudouresque et al. (1990), at least eight species of macroalgae that live in coralligenous communities can be considered endangered: *Chondrymenia lobata*, *Halarachnion ligulatum*, *Halymenia trigona*, *Platoma cyclocolpa*, *Nemastoma dichotomum*, *Ptilophora mediterranea*, *Schizymenia dubyi* and *Laminaria rodriguezii*. However, this list can be greatly extended by adding species such as *Aeodes marginata*, *Sphaerococcus rhizophylloides*, *Schmitzia neapolitana*, *Ptilocladiopsis horrida*, *Microcladia glandulosa*, *Rodriguezella bornetii*, *R. pinnata* and *Lomentaria subdichotoma* (Ballesteros, unpublished data). Most of these species have coralligenous or maërl beds as their only habitats, and seem to be very sensitive to pollution and increased sedimentation rates (Boudouresque et al. 1990), two of the main threats to coralligenous assemblages. The case of *Laminaria rodriguezii* is especially relevant, as this species develops best in rhodolith beds, from where it has almost disappeared due to trawling activities; coralligenous bottoms now constitute its only refuge.

Several animal species in coralligenous habitats are also considered to be at risk (Boudouresque et al. 1991). Although none of them is in danger of extinction, local depletion of some species stocks may occur. Most of the endangered species have great commercial value and this is the main reason for their increased rarity.

Among the anthozoans, red coral (*Corallium rubrum*) is exploited commercially in almost all Mediterranean countries, and its stocks have strongly declined in most areas, particularly in shallow waters (Weinberg 1991). Populations of gorgonians common in coralligenous communities but which lack commercial value, such as *Paramuricea clavata*, *Eunicella cavolini* and *E. singularis*, are pulled out inadvertently by recreational divers (Coma et al. 2004). The black coral, *Gerardia savaglia*, is a very rare species and can be a target for collection by divers, thus making the species even scarcer (Boudouresque et al. 1991).

Some species of molluscs living in coralligenous communities are also threatened. The edible rock-borer bivalve *Lithophaga lithophaga* is considered an endangered species (Boudouresque et al.
1991) despite being extremely abundant. Harvesting by divers is only important in shallow waters and the reason behind calls for the species to be protected is actually an attempt to protect the shallow benthic communities in rocky shores dominated by macroalgae (Russo & Cicogna 1991, Hrs-Brenko et al. 1991), not the coralligenous bottoms themselves. Protection of the two species of fan mussels (*Pinna nobilis* and *P. rudis*) present in the Mediterranean has also been proposed (Boudouresque et al. 1991), because they have been decimated in northern Mediterranean areas by coastline modification and harvesting as souvenirs (Vicente & Moreteau 1991). *P. nobilis* mainly grows in seagrass meadows, and its presence in coralligenous communities is very unusual (Vicente & Moreteau 1991). However, *P. rudis* (*= P. pernula*) is frequently seen in coralligenous habitats, at least in the warmer areas of the western Mediterranean (Ballesteros 1998).

According to Templado (1991), neither of the two species of the genus *Charonia* that occur in the Mediterranean is threatened by extinction. *C. lampas* is rare in the northern Mediterranean but rather common in the southwest, whilst *C. tritonis variegata* has been recorded in the eastern and southwestern Mediterranean. Both species are collected and used for decorative purposes but Templado (1991) argues that indirect anthropogenic pressures (coastline development) are the main reason for its increased rarity, or even local extinction.

The sea urchin *Centrostephanus longispinus* is also considered an endangered species by Boudouresque et al. (1991), despite being a rare species in the northwestern Mediterranean, probably due to biogeographical reasons. No anthropogenic pressure has been proposed to explain its rarity.

The slipper lobster, *Scyllarides latus*, is highly appreciated gastronomically. The high market prices it obtains have stimulated increased fishing pressure, which has led to a dramatic decline in the abundance of this species in several areas of the Mediterranean (Spanier 1991). It is more common in the warmer Mediterranean areas (e.g., eastern Mediterranean, Balearic Islands), and rarest in the colder ones.

The dusky grouper, *Epinephelus marginatus* (*= E. guaza*), is the main target species in spearfishing activities and its abundance has greatly decreased in several Mediterranean areas, mainly in the north (Chauvet 1991). However, immature specimens and juveniles are very abundant in certain areas (e.g., Balearic Islands; Riera et al. 1998) and, therefore, the species is only threatened in those places where there is no regular recruitment (e.g., northwestern Mediterranean). Moreover, the recovery of this species in marine protected areas has repeatedly been reported (Bell 1983, Garcia-Rubies & Zabala 1990, Francour 1994, Coll et al. 1999), as has reproduction (Zabala et al. 1997a,b), suggesting that adequate management can rapidly improve its situation in those areas where stocks continue to decline. Other groupers, such as *E. costae* (*= E. alexandrinus*), *Mycteroperca rubra* and *Polyprion americanus* (Riera et al. 1998; Mayol et al. 2000), are probably in a worse situation, as their population stocks are much lower than those of the dusky grouper.

*Sciaena umbra* and *Umbrina cirrosa* are the two other fish considered as endangered in the review by Boudouresque et al. (1991). Both can live in coralligenous communities, the former being more abundant. Although both species are easily spearfished, *Sciaena umbra* stocks readily recover after fishing prohibition (Garcia-Rubies & Zabala 1990, Francour 1994).

Other species are not included in the list of Mediterranean endangered species by Boudouresque et al. (1991), although according to Mayol et al. (2000) they are exposed to major risk. This is the case of several small sharks inhabiting detritic and coralligenous habitats: *Scyliorhinus stellaris, Mustelus asterias, M. mustelus, Squalus acanthias* and *S. blainvillei*. All these species were very common in fish catches by Balearic Island fishermen at the beginning of the twentieth century, but are now extremely rare. Other species that can thrive in coralligenous communities and which are considered by Mayol et al. (2000) to be endangered are seahorses (mainly *Hippocampus ramulosus*), *Gaidropsarus vulgaris* and some cryptobenthic fishes (*Didogobius splechtnai, Gammogobiussteinitzii*). These are not commercial species and their increased rarity may be related to indirect effects of fishing (such as cascading effects), physical disturbances of trawling or other unknown causes.
Biotic relationships

Spatial interactions, herbivory, carnivory

Biotic relationships, both trophic ones and those related to spatial interactions, are a major force in structuring all ecosystems. In fact, the whole buildup of coralligenous frameworks is affected by the interactions between encrusting corallines and other sessile, invertebrate builders (Figures 16A,B). The final result (that is, what the framework looks like) is not only related to which builder has been the most effective but also to how the borers (from sea urchins to excavating sponges and polychaetes) have subsequently changed the structure. Biotic relationships at this level are, therefore, crucial in building coralligenous assemblages.

Trophic relationships are especially interesting in coralligenous communities because the main organisms are not easily edible. Most of them have skeletons that contribute to structure but which also deter feeding (Zabala & Ballesteros 1989). Others may have chemical defences that make them unpalatable or even toxic (Martí 2002). Most of the largest sessile invertebrates living in coralligenous communities do not feed directly upon other animals from the coralligenous assemblage but rather on the pelagic system. In fact, the largest part of the living biomass in coralligenous assemblages consists of algae and suspension feeders (True 1970, Zabala & Ballesteros 1989), which suggests that herbivory and carnivory are not as important as in other marine Mediterranean environments. The low dynamism of coralligenous habitats (Garrabou et al. 2002) also supports this suggestion.

Figure 16 (See also Colour Figure 16 in the insert.) Spatial interactions are crucial in the buildup of coralligenous assemblages. (A) Mesophyllum alternans overgrows Lithophyllum cabiochae which, in its turn, is epiphytised by the small green alga Halicystis parvula (above) and a tunicate (below); (B) Lithophyllum frondosum overgrows sponge Ircinia oros. Strong prey selection is present in the coralligenous community. (C) Opisthobranch Discodoris atromaculata feeds almost exclusively on sponge Petrosia ficiformis; (D) Opisthobranch Flabellina affinis feeds on hydrozoans of the genus Eudendrium. (Photos by the author.)
However, both herbivory and carnivory are relevant to coralligenous communities. The sea urchin *Sphaerechinus granularis* is a major browser of encrusting corallines (Sartoretto & Francour 1997), and several invertebrates (opisthobranchs, amphipods, copepods) are able to feed on the green alga *Halimeda* (Ros 1978). Examples of carnivores include most of the fishes that thrive in coralligenous communities, as well as most prosobranchs, echinoderms, vagile polychaetes and crustaceans. Although feeding by most animals is not selective, there are some noteworthy examples of animals that have a strong prey selection. These include the well-known cases of the opisthobranch *Discodoris atromaculata*, which feeds on the sponge *Petrosia ficiformis* (Figure 16C), and the other opisthobranch *Flabellina affinis*, which feeds mainly on hydrozoans of the genus *Eudendrium* (Figure 16D) (Ros 1978). Other interesting examples have recently been reported for copepods of the genus *Asterocheres*, which systematically feed on both rhagons and adult sponges by sucking the material produced at the ectosome of the sponge (Mariani & Uriz 2001).

**Chemical ecology**

The production of active substances in benthic organisms plays a major role in structuring benthic communities. Some of these substances act as a defence against consumers (e.g., unpalatable or repellent substances) while others mediate the interactions between species regarding the occupation of space (Martí 2002). Sponges, bryozoans and tunicates are the taxa with the largest number of species producing active substances (Uriz et al. 1991). The lower side of coralligenous blocks, as well as semidark caves and overhangs, exhibits the highest percentage of active species of all the Mediterranean communities sampled by Uriz et al. (1991), suggesting that investment in production of allelochemicals plays an important role in space competition in coralligenous assemblages (Figure 17A).

**Epibiosis, mutualism, commensalism, parasitism**

There are innumerable relationships between species in coralligenous communities that can be described as ‘associations’, and these may or may not involve trophic transfer. Sometimes it is difficult to differentiate between them because the natural history of the species, or the benefits and costs of the components of the association, are unknown or not clearly understood. The purpose here is not to review these associations, nor to mention all those which have been described for coralligenous communities, but to report some examples of epibiosis, mutualism, commensalism and parasitism that can give an idea of the complexity of the coralligenous community with respect to these kinds of relationships.

Epibiosis is a widespread phenomenon in benthic communities and coralligenous assemblages are an excellent example of the different strategies adopted by organisms to cope with this problem (True 1970). Some basibionts tolerate different degrees of epibiosis and even almost complete overgrowth, whilst others have developed antifouling defences to avoid overgrowth. Both types of strategies can be displayed by species from the same zoological group living in coralligenous communities. For example, the ascidians *Microcosmus sabatieri* and *Pyura dura* are usually completely covered by a wide array of epibionts, whilst *Halocynthia papillosa* and *Ciona edwardsi* are always free of overgrowing organisms (Ramos 1991).

Some epibionts are considered to select their hosts, whilst others are not selective. The anthozoan *Parerythropodium coralloides* usually grows over the axes of gorgonians (*Eunicella, Paramuricea clavata*), although it can also grow over other animals and seaweeds, or be attached to rubble or any other kind of substratum (Laubier 1966, Gili 1986). The anthozoan *Parazoanthus*
axinellae prefers sponges of the genus Axinella (mainly *A. damicornis*) (Figure 17B), but it can also grow over other sponges or over rock or encrusting corallines (Gili 1986). The bryozoan *Turbicellepora avicularis* prefers the basal parts of the axes of gorgonians *Paramuricea clavata* and *Eunicella* spp. (Laubier 1966, Zabala 1986).

The number of species able to act as nonselective epibions in coralligenous communities is huge because most of the space is occupied and larvae usually have to settle on living animals or plants. Therefore, almost all sessile species can be epibions (True 1970) (Figure 17C). Gautier (1962), for example, reviewed the epibiosis of bryozoans over bryozoans in coralligenous assemblages, and Nikolic (1960) reported up to 18 species growing over *Hippodiplosia foliacea* in a coralligenous framework in the Adriatic Sea. Of particular interest are the observations by Laubier (1966) on some heterotrichs (Protozoa) of the family Folliculinidae that live close to the zooid mouth of different species of bryozoans or even inside its empty zooids. Laubier (1966) reported up to six species of Folliculinidae living as epibions of bryozoans in the coralligenous communities of Banyuls.

Mutualism has been reported, for example, in the case of the scyphozoan *Nausitoë punctata* and several horny sponges (Uriz et al. 1992b). *Cacospongia scalaris, Dysidea avara* and *D. fragilis* utilize the thecae of *Nausitoë punctata* as a substitute for skeletal fibres, presumably reducing metabolic costs associated with skeleton building. The scyphozoan should thus benefit from greater protection against predation and mechanical disturbance, trophic advantages (inhalant flow carries out small particles susceptible to capture by the scyphozoan), and chemical defence against predators, as the three species of sponges exhibit toxicity (Uriz et al. 1992c).

Commensalism is one of the most common relationships in coralligenous communities. Most relationships are considered as commensalism because they lack unequivocal parasitic features, as in the case of the polychaete *Eunice siciliensis* and the decapods *Alpheus dentipes* and *Typton..."
spongicola, which live as endobionts of the boring sponge Cliona viridis (Laubier 1966). Another well-known example is that occurring between C. viridis and the likewise boring polychaete Dipolydora rogeri (Martin 1996). The curious feature of this association is that it persists even when the sponge is massive and nonexcavating. The feeding activity of the polychaete is favoured by the inhalant flow of the sponge and, moreover, the sponge offers physical protection to the worm when they are not embedded within the calcareous algae. The ability of the worm to manipulate relatively large particles (either to feed or to build its tubes) may favour the filtering activity of the sponge by cleaning the area around the inhalant papillae, thus preventing the collapse of their orifices (Martin & Britayev 1998).

Polychaetes of the genus Haplosyllis are well-known commensalists (Martin & Britayev 1998). Up to 200 specimens of Haplosyllis spongicola have been found in 1 cm² of sponge (Bacescu 1971), and thus sponges merit the description of ‘living hotels’. Another case is the polychaete Haplosyllis depressa chamaeleon, which lives exclusively as a commensal of the sea fan Paramuricea clavata, where it crawls above the living colonies (Laubier 1960, 1966).

The barnacle Acasta spongites lives inside the sponge Ircinia variabilis, as well as other sponges (Laubier 1966, Rützler 1976, Uriz et al. 1992b); it can be considered as a parasite because the cirripede settles into the inhalant oscula of the sponge, rendering it useless. Another barnacle, Pyrgoma anglicum, is quite often found living inside the antozoan Leptopsammnia pravoti (Figure 17D). A further example of parasitism in coralligenous communities is found in the two boring spionid polychaetes Dipolydora armata and Polydora hoplura and the bryozoans Dentiporella sardonica, Porella concinna, Brodiella armata and Schizomavella auriculata (Laubier 1959a, 1959b). The bryozoans are infested throughout their basal layer and polychaetes excavate galleries that reach the surface of the colony or modify the growth form of the bryozoan in such a way that the polychaete tubes are composed of host zooids. In both cases the bryozoan is stimulated to build calcareous formations around the end of the polychaete tubes, facilitating the feeding behaviour of the worm and protecting it (Laubier 1966).

Processes

**Growth and age of coralligenous frameworks**

The mean growth rate of pillars of Mesophyllum alternans (reported as M. lichenoides) in La Ciotat (NW Mediterranean) has been estimated by radiocarbon dating to be 0.19 mm yr⁻¹ (Sartoretto 1994), with a range of 0.11 to 0.26 mm yr⁻¹. Similar values of 0.16 mm yr⁻¹ over the last 640 yr were obtained in a coralligenous block sampled at 15 m in the Natural Reserve of Scandola (Corsica) (Sartoretto et al. 1996).

Ages obtained by radiocarbon dating of coralligenous frameworks situated at depths between 10 and 60 m in the northwestern Mediterranean range from 640 ± 120 yr before the present (BP) to 7760 ± 80 yr BP (Sartoretto et al. 1996). Internal erosion surfaces within the concretions provide evidence of discontinuous development. The accumulation rate of the coralligenous constructions is very low (0.006–0.83 mm yr⁻¹) and oscillates greatly according to depth and time period. The highest accumulation rates (0.20–0.83 mm yr⁻¹) have been recorded for deep coralligenous frameworks and correspond to a period between 8000 and 5000 yrs BP. After 5000 yr BP, the only appreciable accumulation rates (0.11–0.42 mm yr⁻¹) have been recorded for coralligenous frameworks situated in relatively shallow waters (10–35 m depth), whereas the accumulation rates of concretions below a depth of 50 m is almost nil. Thus, the development of these deep coralligenous formations occurred a long time ago, when the depth of the overlying layer of water probably did not exceed 10–15 m, and since their period of settlement (8500–7000 yr BP) the accumulation rate has progressively declined. This decrease in accumulation rates can no doubt be explained by the
stable, but rather unfavourable, environmental conditions resulting from the decrease in irradiance related to increased water depth. It is also worth noting that the coralligenous frameworks below a depth of 30 m in the Marseilles area are today inactive and almost devoid of living coralline algae. In contrast, those present in Corsica at a depth of 50 m and below (down to 65 m) are completely covered by living corallines and are in an active period of growth; this difference must be related to the clear waters present in Corsica.

Carbonate production

Although there are no specific studies on the carbonate production of the coralligenous community on a yearly basis, Canals & Ballesteros (1997) estimated the carbonate production of the phyto-benthos in the coralligenous and other communities from the continental shelf of the Balearic Islands, taking into account the standing crop of calcareous algae and their P/B ratios. The coralligenous community thriving in relatively shallow waters (with *Mesophyllum alternans* and *Halimeda tuna* as dominant algae) was the one with the highest production (around 465 g CaCO₃ m⁻² yr⁻¹). Production of deep water coralligenous concretions dominated by *Lithophyllum cabiochae* was much lower (around 170 g CaCO₃ m⁻² yr⁻¹), but even this rate is much higher than average carbonate production for the Balearic shelf as a whole (100 g CaCO₃ m⁻² yr⁻¹). The contribution of suspension feeders to the total carbonate production of coralligenous communities in the Balearic shelf, as well as in other Mediterranean areas, is practically unknown. However, estimates of the animal carbonate production in deep water (25–50 m depth) rocky bottoms from the Alboran Sea (southwestern Mediterranean), mainly dominated by big suspension feeders with calcareous skeletons (predominantly the coral *Dendrophyllia ramea*, the bryozoans *Pentapora fascialis*, *Smittina cervicornis* and *Myriapora truncata*, and polychaetes such as *Salmacina dysteri*, *Protula* sp. and other serpulids), are very high (around 660 g CaCO₃ m⁻² yr⁻¹) (Cebrian et al. 2000). Although the combination of highly productive calcareous animals in these bottoms is unusual in other Mediterranean areas, this figure can be considered as the upper limit for animal carbonate production in the coralligenous habitat.

Bioerosion

The most active browser in the coralligenous community is the sea urchin *Sphaerechinus granularis*, which accounts for a large part of the total coralligenous erosion. Sartoretto & Francour (1997) calculated an erosion rate ranging between 16 and 210 g CaCO₃ m⁻² yr⁻¹, with higher values in shallow waters and lower values in coralligenous concretions around 50 m depth. The bioerosional role of *Echinus melo* cannot be measured, but it is very low. Among macroborers the spionid polychaetes *Polydora* spp. and the mollusc *Hyatella arctica* are the only macrofauna that colonise experimental blocks after 1 yr of exposure (Sartoretto 1998). In this study, the total erosion caused by annelids and molluscs increases with the number of individuals but does not exceed 5.73 ± 0.77 g CaCO₃ m⁻² at 20 m and 1.50 ± 0.99 g CaCO₃ m⁻² at 60 m after 1 yr. In natural communities macroborers are more abundant (excavating sponges, Sipunculida, perforating molluscs) and their absence in the experimental substrata may be due to their slow growth and to the high spatial and temporal variability of larval recruitment (Kleeman 1973).

The comparative erosion rates produced by the three main types of eroding organisms (browsers, microborers and macroborers) have been estimated by Sartoretto (1996). Sea urchins account for roughly 95% of the total mass of CaCO₃ eroded. Bioerosion by micro- and macroborers accounts for the remaining 5%, that of microborers being very low due to the great depths at which
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coralligenous frameworks develop, and the high sedimentation rates, factors that prevent any significant development of microborers (Sartoretto 1998). Bioerosion by macroborers is probably underestimated because the studies of Sartoretto (1998) do not take into account several organisms that are common in coralligenous communities, in particular the bivalve *Lithophaga lithophaga* and excavating sponges (Sartoretto et al. 1996). The overall range of bioerosion observed in the coralligenous community is between 220 g CaCO₃ m⁻² yr⁻¹ in relatively shallow waters and 20 g CaCO₃ m⁻² yr⁻¹ at a depth of 60 m. This range is in agreement with the structure and age of coralligenous frameworks situated below 50 m depth in the region of Marseilles (Sartoretto 1996), and are at least one order of magnitude below the bioerosion experimentally estimated in coral reefs (Chazottes et al. 1995).

*Sedimentation*

The rugosity of coralligenous frameworks promotes the deposition of particles that take part in the buildup of the coralligenous structure through complex processes of lithification (Marshall 1983). However, high sedimentation rates can be a problem because sediment particles may cover the thalli of the encrusting corallines and screen them against light (Laborel 1961), as well as prevent the recruitment of new plants (Sartoretto 1996). Not all the sediment particles deposited in the coralligenous structure are included in the concretion; indeed, many of them are eliminated by different browsers, while others are resuspended by currents, organisms and gravitation. The rugosity of different coralligenous types is different, as is the capacity for sediment retention. The amount of water movement is also important, and this usually decreases with depth. Studies conducted by Sartoretto (1996) in the Marseilles region conclude that around 9 kg m⁻² yr⁻¹ is retained in the coralligenous concretions situated at a depth of between 30 and 60 m, while the retention in shallow waters is much lower. However, daily sedimentation rates are extremely high in shallow waters (~500 g m⁻²) and much lower in deep waters (between 10 and 35 g m⁻²) (Sartoretto 1996).

The sediment that is finally incorporated into the coralligenous framework by a process of lithification has a large calcareous component of organisms living in the coralligenous community. Major contributors are coralline algae (57%), bryozoans (19%), molluscs (16%), corals and serpulids (3% each) (Laubier 1966).

*Dynamics and seasonality*

The study of dynamics for the whole coralligenous community is a very difficult task and has only very recently been undertaken. Garrabou et al. (2002) used a photographic method to look for changes occurring in two monitored areas of coralligenous concretions covering 310 cm² over a period of 2 yr. The rate of change observed averaged 10% month⁻¹, with very low to nil seasonality. Most of the area (>70%) remained almost constant throughout the 2 yr of monitoring, showing no or few transitions, and this indicates the great persistence of the animals and plants that thrive in coralligenous communities (Figure 18).

Other studies have been conducted with some components or species of coralligenous communities. In fact, most studies dealing with the biology of the main species in coralligenous communities (see next section) have described the effects of seasonality, when this process exists.

In terms of benthic flora, Ballesteros (1991a) described the seasonal cycle of several phytobenthic communities from the northwestern Mediterranean, making a between-community comparison using the same variables as descriptors. The coralligenous community with *Mesophyllum alternans* and *Halimeda tuna* had the lowest seasonality of all the subtidal communities studied, this being almost constant in autumn, winter and spring, but with peak productivity in summer,
During which time there were higher biomass values than for the rest of the year. Piazzi et al. (2004) found significant seasonal differences that were mostly related to the disappearance of many turf species and the decrease in cover of most erect algae, principally foliose and corticated-terete forms, in winter. Although growth of coralline algae is almost constant throughout the year (Garrabou & Ballesteros 2000), *Halimeda* growth occurs mainly in summer (Ballesteros 1991c). In terms of structural changes in the community, two stages can be discerned over an annual cycle: a diversified community stage, with a reduced coverage of *Halimeda* and other soft algae, and a developed community stage, characterized by a high coverage of *Halimeda* (Ballesteros 1991b). The shift from the diversified community stage to the developed community stage takes place through a production phase (early summer). A diversification phase can be distinguished in late autumn, when a sudden fall in *Halimeda* coverage is detected (Ballesteros 1991b) (Figure 19).

Most benthic hydrozoans exhibit a seasonal pattern, with reproduction in spring or autumn and growth from autumn to spring; most of them disappear during the summer, leaving only dormant basal stolons (Boero et al. 1986). Epiphytic hydrozoans on *Halimeda tuna* decline in abundance in summer because of the death of old thalli of *Halimeda*, the growth of new thalli and apical articles on existing thalli, and possibly because of interspecific competition with epiphytic algae (Llobet et al. 1991a).

**Figure 18** (See also Colour Figure 18 in the insert.) Maps of transition intensity resulting from overlay procedures of images from the same plot (310 cm²) along a depth gradient in a vertical wall at the Medes Islands (NE Spain) during 2 yr of sampling. Patch colour denotes number of changes taking place in each patch (see legend). Coralligenous communities (14 and 20 m depth) display much lower transition rates than shallow water communities, indicating high persistence and low rates of change in the animals and plants thriving in the coralligenous communities. (From Garrabou et al. 2002. With permission from Elsevier.)
Anthozoans exhibit a marked seasonality in all activities (Coma et al. 1998a, Garrabou 1999). According to Coma et al. (2002) respiration rates of *Paramuricea clavata*, *Dysidea avara* and *Halocynthia papillosa* vary two- to three-fold across the annual cycle, exhibiting a marked seasonal pattern but showing no daily cycle or significant day-to-day variability within months. The respiration rate of *Paramuricea* (a passive suspension feeder) does not correlate with temperature, but that of *Dysidea* and *Halocynthia* (active suspension feeders) increases with temperature. There is a low rate of new tissue synthesis during summer, together with the contraction of polyps and a low Q₁₀, which explains the low respiration rates of *Paramuricea* observed during the period of highest temperature. These low respiration rates support the hypothesis that energy limitations may underlie summer dormancy in some benthic suspension-feeding taxa in the Mediterranean (Figure 20).

**Figure 19** Although seasonality in structural patterns is not very evident in coralligenous communities, assemblages of the green alga *Halimeda tuna* show high production in summer, higher biomass in autumn, low production in winter and high spatial heterogeneity in spring, going through the two stages of a diversified community (spring) and a developed community (autumn). (From Ballesteros 1991a.)

Activity rhythm in the gorgonian *Paramuricea clavata*, estimated as a percentage of expanded colonies, displays a strong decrease in summer, in conjunction with high water temperatures. This, and other evidence of decreased activity (i.e., growth and reproduction), in *Paramuricea*, as well as in other suspension-feeders, prompted Coma et al. (2002) to describe summer dormancy for many Mediterranean benthic invertebrates. (From Coma et al. 1998a. With permission from Inter-Research.)
There is growing evidence that seasonal patterns of activity and secondary production of suspension feeders in coralligenous assemblages are characterized by aestivation (Coma et al. 2000). Several types of resting and resistance periods have been observed in several colonial ascidians in the warm season (Turon 1992, Turon & Becerro 1992). In the case of *Polysyncraton lacazei* the surface of the colonies is covered by a glassy pellicle and the siphonal apertures are sealed. This state is interpreted as a rejuvenative phenomenon that extends the life span of the zooids (Turon 1992). Some sponges also go through a resting, nonfeeding period with cellular restructuring, mainly in summer. For example, some specimens of *Crambe crambe* appear to be covered by a glassy cuticle, obliterating the oscula and ostia after reproduction, from the end of August until the end of October (Turon et al. 1999). These authors suggest that these resting stages develop not only in response to remodelling following reproduction, but also as an effect of water temperature abnormalities.

The decapod fauna also displays a certain seasonality (García-Raso & Fernández Muñoz 1987), due to the intense recruitment of several species in late summer, and a progressive decrease in the density of individuals and an increase in their size from October to June.

The fish fauna of coralligenous communities is also affected by seasonality (Garcia-Rubies 1997), although its effect is of very minor importance. The number of species in fish counts along 50 m-long visual transects of the coralligenous bottoms around the Medes Islands slightly decreases in winter, and most fishes seem to be less active than in summer.

**Functioning of outstanding and key species**

Several studies of coralligenous concretions are devoted to species that are particularly abundant, are architecturally important or are economically valuable. A compilation of the major knowledge of these species is presented here.

**Coralline algae**

Growth dynamics of two important coralligenous builders in the northwestern Mediterranean, *Mesophyllum alternans* and *Lithophyllum frondosum*, were studied in the bioconcretions of the Medes Islands marine reserve, in a steep wall situated at a depth of between 15 and 30 m (Garrabou & Ballesteros 2000). Growth rates ranged from 0.16 month\(^{-1}\) for *Mesophyllum alternans* to 0.09 month\(^{-1}\) for *Lithophyllum frondosum*, with shrinkage rates being 0.09 and 0.04 month\(^{-1}\), respectively. These growth rates are more than one order of magnitude lower than those reported for other Mediterranean and tropical coralline species, but similar to reports for crustose corallines in Arctic and temperate waters. No seasonal pattern in growth or shrinkage was found for either species, although seasonality in conceptacle occurrence was detected in *Lithophyllum frondosum*, with a high interannual variability. *Mesophyllum alternans* thalli frequently underwent fissions and fusions (almost one event during the 2-yr monitoring period for 50% of monitored plants), while they were rarely observed in *Lithophyllum frondosum*. These differences in growth, shrinkage, and fission and fusion events are interpreted as different growth strategies. *Mesophyllum alternans* has a more opportunistic strategy, growing faster and gaining area more rapidly, although it also loses area at higher rates. *Lithophyllum frondosum* has a more conservative strategy and is more effective in maintaining the area acquired through its reduced growth rate (Garrabou & Ballesteros 2000).

**Halimeda tuna**

Growth and production of a *Halimeda tuna* population from a coralligenous community (18 m depth) in the northwestern Mediterranean was studied by Ballesteros (1991c). The production of new segments changed seasonally, being maximal in summer and minimal in winter (Figure 21),
and this suggests that growth is mainly related to temperature and irradiance. The loss of segments seemed to be related to physical disturbances (storms) and herbivory. Annual production of *H. tuna* was estimated at 680 g dw m$^{-2}$, equivalent to 114 g organic C m$^{-2}$ yr$^{-1}$ and to 314 g CaCO$_3$ m$^{-2}$ yr$^{-1}$; the yearly P/B ratio was 1.87 yr$^{-1}$. The epiphytic assemblage growing on the segments of *H. tuna* also displayed high seasonality, with a maximum biomass and species richness in early summer. Values of growth and production reported in Ballesteros (1991c) emphasize the importance of *H. tuna* as a producer both of organic matter and calcium carbonate in the coralligenous habitat. In fact, available data suggest that calcium carbonate production by *Halimeda* in shallow coralligenous concretions is similar to that of coralline algae (Canals et al. 1988).

**Porifera**

Garrabou & Zabala (2001) studied the growth dynamics of four demosponges (*Crambe crambe* and *Hemimycale columella* from a 'precoralligenous' community, and *Oscarella lobularis* and *Chondrosia reniformis* from a coralligenous community in the Medes Islands), and reported relatively slow growth dynamics with low growth and shrinkage rates. The coralligenous species had an average relative growth rate of 0.15 month$^{-1}$ (*Oscarella*) and 0.022 month$^{-1}$ (*Chondrosia*), with shrinkage rates of 0.12 and 0.017 month$^{-1}$, respectively. Interspecific differences in growth, shrinkage, division and fusion rates were interpreted as evidence of distinct biological strategies aimed at persistence and the occupation of substratum. *Chondrosia reniformis* is conservative, with slow growth but great resistance to damage. *Crambe crambe* seems to enhance its rate of space occupation by a high division rate. *Hemimycale columella* grows quickly and shrinks at low rates, thus spreading rapidly over the substratum. *Oscarella lobularis* grows and shrinks rapidly, showing great overall growth.

*Dysidea avara*, a common sponge in coralligenous communities (Uriz et al. 1992a) obtained 85% of its ingested carbon from the fraction <5 μm (mostly procaryotes and pico- and nanoplankton) and 15% from the fraction >5 μm (mostly phytoplankton) (Ribes et al. 1999b). However, the partial contributions of the different groups varied seasonally, in accordance with the planktonic composition of the water column. During winter, phytoplankton was an important component of the total uptake (26%), whereas during the rest of the year it contributed <7% of the total uptake. This trophic plasticity may represent an advantage for the species because it attenuates the effects of

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**Figure 21** Seasonal changes in segment production and loss for a population of the green alga *Halimeda tuna* at 18 m depth in a coralligenous wall off Tossa de Mar (NE Spain). (From Ballesteros 1991c. With permission from Walter de Gruyter GmbH & Co. KG.)
seasonal fluctuations in the planktonic community. Moreover, the water transport rates of 63 ml g−1 min−1 and the high clearance rates for particles measuring 4 μm or less observed in *D. avara* (Turon et al. 1997) point to a significant grazing impact of sponges on the picoplankton in communities like the coralligenous, where *D. avara* and other massive sponges are abundant (Laubier 1966; Bibiloni et al. 1984).

**Hydrozoa**

The hydrozoan *Orthopyxis crenata*, a common hydroid growing on the alga *Halimeda tuna*, was most abundant from November to April, when three cohorts were identified (Llobet et al. 1991b). During the rest of the year only two cohorts were identifiable, except in June and July when there was only one (juvenile) cohort. Reproduction took place from late October to early December, with recruitment occurring at the same time. In winter, colony sizes quintupled and tripled every 15 days, living a maximum of 6 wk. The maximum abundance of hydroids in winter seems to be related to increased food availability, a decrease in competence by epiphytic algae and a decline in the turnover rate of *Halimeda* (Ballesteros 1991c). The strategy of *Orthopyxis crenata* and other hydroids is completely different to that of long-living anthozoans that are also common in coralligenous communities; the colonies survive for only a small number of weeks but asexual reproduction by the creeping stolons ensures colony survival beyond the life of *Halimeda* thalli, and perhaps indefinitely.

Coma et al. (1992) studied the life cycle of two similar species of hydrozoans living over the thalli of *H. tuna* and also found that survival should be very long for each colony, due to the active asexual reproduction (by stolonisation in *Halecium petrosum*; by planktonic propagules in *H. pusillum*) that occurs throughout most of the year. Maximum life span of colonies was estimated to be only 8 wk, mean colony sizes increasing between three- and six-fold over consecutive 2-wk periods.

**Corallium rubrum**

Red coral (*Corallium rubrum*) is typically associated with the animal dominated communities growing in dim light conditions and which characterize smaller cavities, vertical cliffs and overhangs present in coralligenous concretions. Although it is predominantly found in the western basin, it is also present in some areas of the eastern basin and the African-Atlantic coast (Zibrowius et al. 1984, Chintiroglou et al. 1989).

Harvesting is the major source of disturbance in red coral populations (Santangelo et al. 1993, Santangelo & Abbiati 2001), although large-scale mortalities have also been documented (Arnoux et al. 1992, Garrabou et al. 2001). Recreational diving seems to have a limited impact on populations but the potential risks of poaching and mechanical disturbance will increase in the near future with the predictable increase of diving (Garrabou & Harmelin 2002).

According to Garrabou & Harmelin (2002), red coral has a high survivorship, with 60% of colonies reaching 22 years of age. Mortality is higher in juveniles, but partial mortality of colonies is higher in old colonies (Garrabou & Harmelin 2002). Age at first reproduction is, on average, between 7 and 10 yr, although the reproductive effort, i.e., the percentage of gravid polyps per colony, is higher in older colonies (Torrents et al. 2005). Male gonads develop within 1 yr, whereas the maturation of female gonads takes 2 yr (Vighi 1972). Planulae emission seems to be related to water temperature and lasts from May–October, according to each geographical area (Lacaze-Duthiers 1864, Lo Bianco 1909, Cerruti 1921, Vighi 1972, Santangelo et al. 2003). There is indirect
evidence to suggest that the larval dispersal capability of red coral is reduced (Weinberg 1979, Abbiati et al. 1993), and it seems that genetic exchange between populations is also limited, promoting the presence of several discrete and distinct populations in the Mediterranean (Abbiati et al. 1993).

Recruitment, when studied in experimental panels, is usually high at the beginning, subsequently decreasing (Garrabou & Harmelin 2002, Bramanti et al. 2005). However, differences in the results obtained by Garrabou & Harmelin (2002) and Bramanti et al. (2005) suggest there is high population variability in recruitment and colonization rates.

Growth rates for red coral estimated by petrographic methods range from 1.57–0.91 mm yr\(^{-1}\) for basal diameter (García-Rodríguez & Massó 1986, Abbiati et al. 1992), but estimates decrease to 0.62 mm yr\(^{-1}\) in mid-term (4 yr) observations (Cerrano et al. 1999, Bramanti et al. 2005) and to 0.24 mm yr\(^{-1}\) in long-term (22 yr) studies (Garrabou & Harmelin 2002). A new technique for aging red coral developed by Marschal et al. (2005) suggests mean growth rates of around 0.35 mm yr\(^{-1}\), in close agreement with long-term observations. Growth rates in colony height have been estimated to be around 1.8 mm yr\(^{-1}\) (Garrabou & Harmelin 2002, Bramanti et al. 2005). The average branching rate for each colony is 3.4 branches in 22 yr (Garrabou & Harmelin 2002). Harvested populations show about two-fold lower values on average, and up to four-fold lower values in colony size compared with nonharvested populations. Garrabou & Harmelin (2002) provide indisputable data on the longevity of colonies and the parsimonious population dynamics of *C. rubrum*. Current populations have shown a dramatic shift in their size structure, characterized by the absence of large colonies (Figure 22). Full recovery of shallow-water harvested populations may take several decades or even centuries (Garrabou & Harmelin, 2002).

No sign of predation has been observed in monitored colonies of *C. rubrum* (Garrabou & Harmelin 2002). In fact, predation appears to play a minor role in sessile, invertebrate-dominated communities as a whole (Garrabou et al. 2002).

**Figure 22** (See also Colour Figure 22 in the insert.) A red coral colony (age unknown) from a pristine site (Cap Creus, 35 m) collected in 1962 (A), and a 28-year-old colony from an experimental panel (Riou Caramassaigne, 62 m) (B). (Photo and data courtesy of J.G. Harmelin.)
Growth of *Paramuricea clavata* has been monitored photographically over a 2-yr period (Coma et al. 1998b). Based on growth-rate data, the largest colonies in the population (55 cm) were around 31 years old (Figure 23). On average, net production for all colonies was 75% of gross production. Gross production was 4.4 g ash free dry weight (afdw) m$^{-2}$ yr$^{-1}$, while the P/B value was 0.11 yr$^{-1}$ and the turnover time was 9 yr (Coma et al. 1998a,b). Mistri & Ceccherelli (1994), in a study of *P. clavata* in the Straits of Messina (Italy), estimated a production of 3 g afdw m$^{-2}$ yr$^{-1}$, a P/B value of 0.13 yr$^{-1}$ and a turnover rate of 7.5 yr. In *Eunicella cavolinii*, Weinbauer & Velimirov (1995a,b) have estimated a production of 0.3–7.4 g afdw m$^{-2}$ yr$^{-1}$, a P/B around 0.24 and 0.32 yr$^{-1}$, and a turnover rate ranging from 3–4 years.

Mean increase in maximum height for *Paramuricea clavata* ranges from 1.8–2.7 cm yr$^{-1}$ (Weinberg & Weinberg 1979, Mistri & Ceccherelli 1994, Coma et al. 1998a). Similar values have been obtained for *Eunicella singularis* (2.2 cm yr$^{-1}$; Weinberg & Weinberg 1979). Growth is lower in *E. cavolinii* (0.85–1.14 cm yr$^{-1}$; Velimirov 1975, Weinbauer & Velimirov 1995a).

Seasonality of growth in *Paramuricea clavata* requires long monitoring periods in order to be accurately detected, but available data suggest there is a high growth period in spring (Figure 24), this being consistent with the seasonal fluctuation in food sources (Coma et al. 1998b).

The minimum age at first reproduction in *P. clavata* has been estimated to be around 7–13 yr on average (Coma et al. 1995a). Fecundity levels increase with colony size. Oogenesis in *P. clavata* lasts for 13–18 months and culminates with the release of mature eggs in June–July; reproduction is synchronous each year and, as well as coinciding with increasing water temperature, it is correlated with the lunar cycle (Coma et al. 1995a). Spawned eggs adhere to the outer surfaces of female colonies through the action of a mucous coating. Embryogeny and final maturation take place among the polyps. On leaving the surface of the colonies, larvae immediately settle on the surrounding substratum. Maintenance of the population is based on sexual reproduction (Coma et al. 1995b).

Zooplankton (nauplii, copepod eggs, other invertebrate eggs, calanoid copepods) accounts for an important share of the diet. Peak prey capture levels are recorded in spring and at the end of autumn; they fall off substantially in summer, when the proportion of colonies with contracted polyps is very high. The prey capture rate extrapolated to an annual cycle suggests that gorgonians play an important role in the flow of energy from plankton to the benthos; estimates from *P. clavata* populations situated in the Medes Islands indicate that this species can remove the equivalent of between 12 and 85 mg C m$^{-2}$ day$^{-1}$ from the zooplankton (Coma et al. 1994).
However, *P. clavata* has a broad and heterogenous diet that ranges from nano-eukaryotes (3.8 μm) to copepods (700 μm), and includes prey as diverse as ciliates, dinoflagellates, diatoms and suspended detrital organic matter (Ribes et al. 1999c). Carbon of detrital origin accounts for roughly 48% of the total ingested carbon and shows a marked seasonal pattern, in which winter and spring are the seasons with the highest ingestion rates. The amount of carbon removed from the surrounding water is equivalent to 2.7 mg C m⁻² day⁻¹ from the living POC (including nano-eukaryotes, diatoms, ciliates and dinoflagellates) and 28.7 mg C m⁻² day⁻¹ from the detrital POC. No significant capture of dissolved organic matter or picoplankton has been observed. Ribes et al. (1999c) give an estimate of the partitioning of food sources that cover the energy needs of *P. clavata*, assuming data on ingestion rates observed in incubation chambers corrected by the effect of flow speed obtained from the literature. According to these authors, zooplankton and detrital POC make a similar contribution (about 48% each), with the living POC accounting for the remaining 4%, a figure that can probably be extrapolated to other gorgonians.

**Alcyonaria**

The alcyonarian *Alcyonium acaule* has a very slow growth, which is almost undetectable over a 2-yr period (Garrabou 1999). Recruitment is very low and occurs in autumn, this being the only method of population maintenance because there is no asexual reproduction. Mortality rates average 12.7% yr⁻¹, with much higher mortalities in small colonies. Contracted colonies are much more frequent in summer than in any other season (up to 80%) (Garrabou 1999, Rossi 2001). Colonies of *A. acaule* are usually aggregated due to the retention of eggs by the mucous strings, implying a short-range dispersal for larvae and settlement near the parental colonies (Garrabou 1999).

**Zoantharia**

Growth and occupation of space of *Parazoanthus axinellae* in the coralligenous communities of the Medes Islands (northwestern Mediterranean) have been studied by Garrabou (1999). It has moderate growth dynamics (relative growth rate of 0.11 month⁻¹ and a shrinkage rate of 0.09 month⁻¹) with nonsignificant differences over time, although growth rates peak during the summer to autumn period. Fission is common, 29% of monitored colonies undergoing at least one fission.
in 2 yr. Fusion was less frequent, with only 8% of colonies undergoing fusion in 2 yr. Mortality rates average 9% yr⁻¹. The occupation of space in *P. axinellae* seems to be based on spreading over the substrata at moderate rates, either by somatic growth or by fission. Most of the colonies (60%) were contracted in summer and mid autumn (Garrabou 1999, Rossi 2001).

*Tunicata*

Colonies of *Cystodites dellechiajei*, a very common ascidian in coralligenous communities (Ramos 1991), exhibited no or only restricted growth over a 2-yr period in a vertical wall at a depth of 10–12 m in the northwestern Mediterranean (Turon & Becerro 1992), although one of eleven colonies was able to grow actively. According to the authors’ observations, only the individuals present in the most shaded microenvironments displayed active growth. Gonads were present in the population for most of the year. This species shows a high relative biomass per unit area, low growth rates, high survival values (Turon & Becerro 1992) and the presence of chemical defences (Uriz et al. 1991), which would seem to indicate a very conservative life strategy.

Two solitary ascidian species common in coralligenous assemblages spawn in late summer and early autumn. Gamete release occurred after the period of highest temperature (September–October) in *Halocynthia papillosa*, while for *Microcosmus sabatieri* it occurred in October–November (Becerro & Turon 1992). In order to explain the surprising fact of spawning after summer, a period of temperature and food limitation, Ribes et al. (1998) studied the natural diet and prey capture of the ascidian *Halocynthia papillosa* across an annual cycle. The natural diet included detrital organic matter, bacteria, *Prochlorococcus* and *Synechococcus*, protozoans and phytoplankton, with a mean size ranging from <1 μm–70 μm. One specimen of *Halocynthia papillosa* weighing 0.25 g afdw was estimated to ingest an annual mean of 1305 μg C g afdw⁻¹ h⁻¹. Carbon from detrital origin accounted for 92% of the total ingested carbon, while live carbon accounted for only 8%. Ingestion rates showed a marked seasonal pattern, with the highest ingestion of detrital particles in spring and the highest values of ingestion of live particles in summer and autumn. Ribes et al. (1998) hypothesise that live particles are of more significance in the species’ diet than are particles of detrital origin, because the seasonal variation of ingested nitrogen from live particles explained 91% of the gonadal development variance for the year. Thus, living sestonic organisms, rather than detrital carbon, may be an essential source of nitrogen and other nutrients necessary for growth and reproduction in *H. papillosa*.

**Disturbances**

*Large-scale events*

Several episodes of suspension feeder mortality have been detected in the northwestern Mediterranean (Rivoire 1991, Bavestrello et al. 1994, Cerrano et al. 2000, Perez et al. 2000, Garrabou et al. 2001). Here the existing data on the last large-scale mortality of suspension feeders that affected shallow water assemblages (10–40 m depth) eastwards from Marseilles and in some other areas of the central-western Mediterranean (Minorca; Ballesteros, unpublished data) is reported (Figure 25A,B).

Owing to climatic and hydrographic anomalies in the Ligurian Sea, the characteristic summer conditions of reduced resources, high water column stability and high temperatures (normally during July and August) lasted much longer than usual in the summer of 1999. This coincided with a mass mortality of benthic suspension feeders over several hundred kilometres, affecting coralligenous communities situated at a depth of <40 m (Figure 26), where the temperature anomaly lasted for over a month (Perez et al. 2000, Romano et al. 2000). The accumulated density decrease in
Paramuricea clavata colonies 4 yr after the mass mortality accounts for around half the initial population at the Port-Cros National Park (France) (Linares et al. 2005). Red coral populations thriving above a depth of 30 m were also affected (Garrabou et al. 2001). This large-scale mortality, together with other small-scale mass mortalities (Cerrano et al. 2000) recorded during the past decade in the Mediterranean, may be related to seawater temperature increase and global warming. Some suspension feeders might be able to withstand the normal duration of adverse summer conditions but not an anomalous prolongation of these conditions (Coma et al. 2000; Coma & Ribes 2003), resulting from an energy shortage of suspension feeders related to low food availability in summer.

If mass mortalities are indeed related to the global warming trend, such events might occur again and become more frequent, which would cause profound changes in the specific composition and structure of coralligenous communities. In fact, P. clavata, the suspension feeder most drastically affected in the summer 1999 mortality (Perez et al. 2000), is completely absent above a depth

Figure 25 (See also Colour Figure 25 in the insert.) Disturbances in coralligenous communities. (A) Mortality affecting the gorgonian Paramuricea clavata (Port-Cros, France, autumn 1999); (B) Dense carpets of alien alga Womersleyella setacea cover coralligenous assemblages; the gorgonian Eunicella singularis is also affected by previous partial mortality that occurred in summer 1999 (Minorca, Balearic Islands, summer 2000); (C) Filamentous alien alga Womersleyella setacea invades coralligenous rims dominated by Mesophyllum alternans in Cabrera (Balearic Islands, autumn 1999); (D) Partial mortality and overgrowth by filamentous algae affecting gorgonian Eunicella singularis (Minorca, Balearic Islands, summer 2000). (Photos by the author.)
of 40 m in the warm central Mediterranean waters of the Balearic Islands (Ballesteros, unpublished data), and this may be related to the longer duration of summer conditions in this area. However, the ultimate cause of these mortalities remains unclear, because the temperature anomaly can only have caused physiological stress which, in turn, has triggered the development of some pathogenic agent that would otherwise have remained nonvirulent.

Degradation by waste water

Hong (1980) studied the effects of waste water along three stations situated in a gradient of multisource pollution in the Gulf of Fos (Marseilles), and in an unpolluted reference zone. Biodiversity decreased from the reference station (310 species) to the most polluted zone (214 species), and mainly affected bryozoans, crustaceans and echinoderms; molluscs and polychaetes were largely unaffected. The number of individuals also decreased with increased pollution, as did the biomass of sponges and bryozoans, and the diversity of invertebrates. However, the density of sipunculids as well as the relative abundance of species with a wide ecological distribution was enhanced by pollution (Hong 1980, 1983).

The abundances of the species responsible for accretion and those living in the coralligenous community decrease with the pollution gradient, both in terms of number and density of individuals. There are few data concerning the impact of various pollutants on the growth of coralline algae (Littler 1976), although it is known that orthophosphate ions inhibit calcification (Simkiss 1964). However, Hong (1980) observed that with increased pollution large thalli of *Mesophyllum alternans* are replaced by Peyssonneliaceae, which have a much lower building capacity (Sartoretto 1996). Moreover, the species that act as bioeroders are more abundant in the polluted areas (Hong 1980). Thus, all the available evidence suggests that pollution accelerates the destruction of coralligenous assemblages and inhibits building activity.

Cormaci et al. (1985) studied the deep water phytobenthic communities developing over coralligenous concretions in the Gulf of Augusta, a site that is heavily polluted by both urban and industrial wastewater. Water turbidity seems to be the main factor causing degradation and homogenisation of the phytobenthos. There is a slight decrease in the number of species (26 algal species sample$^{-1}$) when compared with similar sites and depths of unpolluted areas (30–38 algal species sample$^{-1}$) (Furnari et al. 1977, Battiato et al. 1979).
Degradation by fishing

Trawling is probably the most destructive fishing method and is causing degradation of large areas of coralligenous concretions (Boudouresque et al. 1990). Trawling not only causes direct physical damage by breaking down the coralligenous structure and rolling the coralligenous blocks but also negatively affects photosynthetic production of encrusting and erect algae by increasing turbidity and sedimentation rates when applied to adjacent sedimentary bottoms (Palanques et al. 2001).

Special trawling to collect precious red coral with what is known as the ‘Italian Bar’ or ‘St. Andrew Cross’ is highly destructive. Ortiz et al. (1986) reported the capture of up to 50 kg of benthic fauna (mainly gorgonians) in order to collect 15 kg of living red coral in the Alboran Sea. The abovementioned device is so effective at destroying the sea bed that it has also been used for scientific studies of fauna associated with red coral (Templado et al. 1986, Maldonado 1992).

Both traditional and recreational fishing also have an effect on coralligenous communities, although they mainly affect the target species. Fishing leads to a significant decrease in mean specific number of fish species, producing changes in the composition of the community (Bell 1983, Garcia-Rubies & Zabala 1990). This effect is due not only to the nearly total absence of some fishes, demonstrated in two species (Epinephelus marginatus and Sciaena umbra) that are extremely vulnerable to spear-fishing, but also to the notable scarcity of other species (Garcia-Rubies 1999). However, depth acts as a protective factor by limiting the effects of fishing, given the inherent difficulty in locating from the surface the coralligenous bottoms that are isolated from the coast (Garcia-Rubies 1999). No cascading effects through overfishing have so far been detected in coralligenous communities, as they have been in shallow rocky bottoms (Sala et al. 1998), although they may well exist because both densities and sizes of fishes and lobsters have been greatly modified over the last 100 yr. Nevertheless, populations of groupers and other vulnerable fishes rapidly recover after fishing is prohibited (Harmelin 1991, Coll et al. 1999, Harmelin & Robert 2001) and readily exhibit normal socio-behavioural patterns and reproductive success (Zabala et al. 1997a,b).

Degradation by the activity of divers

The coralligenous community is one of the most popular sites for recreational diving in the Mediterranean (Boudouresque 2004b) due to its great variety of life and great visual appeal (Harmelin 1993). Some studies have detected the direct impact of divers on the largest invertebrates of the coralligenous community. Sala et al. (1996) found that the large and fragile calcareous bryozoan Pentapora fascialis was present at all levels of exposure (from overhangs to epibiotic) in locations where diving was not allowed, whereas colonies were largely restricted to cryptic positions at diving locations in the Medes Islands marine reserve. Density, colony diameter and colony height were also significantly lower at frequented than at unfrequented sites. Densities of colonies of the bryozoan P. fascialis showed a significant decrease (50% in 1 yr) after a diving site was opened in the marine protected area of the Medes Islands (Garrabou et al. 1998). The impact was greater on boulders covered by coralligenous concretions than on vertical walls, probably as a result of the protection provided by the dense canopy of the gorgonian Paramuricea clavata. However, these high levels of recreational scuba diving (e.g., >1000 visits site−1 yr−1) also appear to be greatly modifying the natural demographic parameters of P. clavata in the Medes Islands marine reserve and adjacent sites by means of a three-fold increase in adult mortality (Coma et al. 2004). This increase in adult mortality is due to toppling by divers, because annual mortality induced by overgrowth is almost similar in dived and undived sites, whereas annual mortality by toppling in high visitation areas ranges from 4.9–6.9% (1.5% in rarely visited sites). Nonintentional breaking of P. clavata has also been reported by Harmelin & Marinopoulos (1994) from the coralligenous
communities in Port-Cros National Park (France). Anchoring of boats seems also to have a negative impact on coralligenous assemblages, although there are no studies where this has been adequately assessed.

Garrabou et al. (1998) conclude that abrasion by divers may affect other organisms of the coralligenous community because it contains many sessile, long-lived organisms with fragile skeletons and slow growth rates that make them very prone to disturbance by toppling. They suggest that diving might lead the coralligenous community to be dominated by erosion-resistant species, such as encrusting and massive organisms, rather than erect, articulate and foliose species. Therefore, human activities may affect the coralligenous community as a whole. Unfortunately, the paucity of data on turnover rates for most organisms thriving on coralligenous assemblages does not allow a quantitative estimate to be made of diver carrying capacity (Sala et al. 1996).

**Invasive species**

Some species introduced into the Mediterranean have become invasive (Boudouresque & Ribera 1994, Boudouresque & Verlaque 2002) and a number of them can thrive in, or are more or less adapted to, the coralligenous habitat. Currently, only introduced algal species are threatening the coralligenous community and then only in some areas of the Mediterranean.

Probably the most dangerous alien species for the coralligenous community is the small red alga *Womersleyella (Polysiphonia) setacea*, which is currently distributed along most of the Mediterranean basin (see Athanasiadis 1997). This species grows abundantly in coralligenous (and other sublittoral) communities (Figure 25C,D), forming a dense carpet, 1–2 cm thick, over the encrusting corallines that constitute the concretion (*Mesophyllum alternans*, *Lithophyllum cabiochae*, and others) (Ballesteros 2004). The carpet of *Womersleyella setacea* undoubtedly decreases light availability to the encrusting corallines (avoiding or reducing photosynthesis and growth of these algae), increases sediment trapping (Airoldi et al. 1995), excludes other macroalgae by overgrowth and pre-emption (Piazzi et al. 2002), and inhibits recruitment of corallines and other algal and animal species inhabiting the coralligenous community (Ballesteros et al. 1998). This alga is also very successful at establishing itself and persisting from year to year (Airoldi 1998). It therefore may cause enormous damage to the entire coralligenous community. Indeed, the species richness found in sites invaded by *W. setacea* is lower than that observed in noncolonised sites (Piazzi et al. 2002).

Another alien turf alga that is able to grow in deep waters is *Acrothamnion preissii* which, nevertheless, has been mainly reported from maërl beds (Ferrer et al. 1994) and the rhizomes of the seagrass *Posidonia oceanica* (Piazzi et al. 1996). Although present in the coralligenous community of the Balearic Islands (Ballesteros, personal observation), it is never dominant in this environment and always grows together with *Womersleyella setacea*.

*Caulerpa taxifolia* is another species that can threaten the coralligenous community. Although mainly found in relatively shallow waters (Meinesz & Hesse 1991), it has been recorded down to a depth of 99 m (Belsher & Meinesz 1995) and in some areas, such as Cap Martin (France), it has totally invaded the coralligenous community (Meinesz 1999). *C. racemosa var. cylindracea*, another invasive species that is quickly spreading in the Mediterranean (Piazzi et al. 2005), is also able to grow in deep waters where coralligenous assemblages develop (down to a depth of 55 m in the Balearic Islands; Ballesteros 2004) but no information about its impact on the coralligenous community is available.

Two other species that have been reported to act as invaders in the Mediterranean are *Asparagopsis taxiformis* (Ballesteros & Rodríguez-Prieto 1996) and *Lophocladia lallemandii* (Patzner 1998). These two species are becoming increasingly abundant both in shallow bottoms and deep waters around the Balearic Islands. They have been found to a depth of 65 m on coralligenous bottoms with or without *Womersleyella setacea* (Ballesteros, personal observation).
Conclusions regarding current knowledge of coralligenous communities

The coralligenous habitat, as described here, is a typical Mediterranean underwater landscape that basically comprises coralline algae growing in dim light conditions and in relatively calm waters. Although it usually develops in the circalittoral zone it can also thrive in the lower infralittoral zone if irradiance levels are low enough to allow the growth of the encrusting corallines.

Most of the available data come from studies conducted in the western Mediterranean. Almost nothing is known about eastern Mediterranean coralligenous habitats, and this may be related to both the greater depth where the coralligenous habitat develops in this area (usually beyond the normal limit of scientific scuba diving) and to the lack of traditional marine research institutes.

The main distribution of coralligenous communities has been well documented on a large scale: it is common all around the Mediterranean coasts, with the possible exception of those of Lebanon and Israel.

Knowledge about environmental factors is rather poor because the coralligenous habitat is a highly heterogenous system and environmental variables can differ greatly on both a geographical and a microscale. However, even though more data are necessary, basic knowledge of light (irradiance), temperatures, nutrient concentrations and hydrodynamism is available. Irradiance seems to be the most critical factor for the development of coralligenous frameworks. Available data suggest that light levels must range between 1.3 MJ m$^{-2}$ yr$^{-1}$ and 50–100 MJ m$^{-2}$ yr$^{-1}$, that is, between 0.05% and 3% of the surface irradiance.

Two main morphologies can be distinguished: banks, which are built over more or less horizontal substrata, and rims, which develop in the outer parts of marine caves and vertical cliffs.

The coralligenous habitat includes several assemblages due to its great heterogeneity. Algal assemblages develop in open waters and are dominated by several species of encrusting red algae. *Mesophyllum alternans* dominates in relatively shallow waters while *Lithophyllum frondosum*, *L. cabiochae* and *Neogoniolithon mamillosum* are more abundant in deep waters. Two main algal assemblages have been distinguished. Shallow-water assemblages are rich in species of green algae, while deep-water assemblages have a poorer algal flora, with some encrusting and foliaceous red algae. Animal assemblages differ greatly among sites and geographical areas. In open areas, mixed with algae, cnidarians dominate the assemblage (mainly gorgonians) in relatively eutrophic areas, while in more oligotrophic waters sponges and bryozoans dominate. On overhangs and in large cavities the communities of suspension feeders are dominated by anthozoans, sponges and bryozoans. Some research has studied algal and animal builders, as well as bioeroders. However, biomass data for the species composing the assemblages are very scarce.

A considerable amount of research has been done on the biodiversity of coralligenous frameworks. Much more data could probably be gathered by taking into account floristic and faunistic studies, as well as monographs from the different groups that contain comments on the ecological distribution of species. A first estimate on the number of species thriving in coralligenous communities is around 1666 (315 algae, 1241 invertebrates and 110 fishes). Studies dealing with the coralligenous communities of certain areas give the number of species as ranging between 500 and 700 species of marine invertebrates. Also important is the very high density of vagile fauna that inhabit coralligenous outcrops, which can reach >3 invertebrates g$^{-1}$ of coralligenous concretion and, for example, a density of >1 polychaete worm cm$^{-2}$.

Some endangered Mediterranean species live in the coralligenous habitat, although none is exclusive to this environment.

As its diversity is so great, the coralligenous habitat reveals an intense connectivity among its inhabitants. Space competition is strong because the space is completely saturated by organisms, and epibiosis is extremely frequent. Alellochemicals must play an important role in space competition
because coralligenous communities exhibit a very high percentage of chemically active species. Trophic relationships are also strong in coralligenous communities, particularly among vagile species because most of the sessile invertebrates have skeletons that deter feeding. Several examples of mutualism, commensalism and parasitism have been reported. Growth of coralligenous accretions, carbonate production, and bioerosion and sedimentation rates have merited very few studies, although those published have presented very valuable data. They indicate (1) very low accumulation rates which are related to water depth and light availability; (2) the important source of carbonate for the continental shelf represented by coralligenous buildups; (3) relatively low bioerosion rates, at least in deep waters where algal growth is the lowest; and (4) relatively high sedimentation rates. Accretion rates of up to 0.83 mm yr$^{-1}$, carbonate production (vegetal and animal) of up to 1000 g CaCO$_3$ m$^{-2}$ yr$^{-1}$, and values for bioerosion of up to 220 g CaCO$_3$ m$^{-2}$ year$^{-1}$ have been reported. These values are always higher in shallow than in deep waters.

Large animals and plants of coralligenous assemblages are highly persistent, and show low to nil seasonality in terms of space occupation. Most of the area covered by a coralligenous community remains unchanged after, for example, 2 yr of monitoring. However, growth pulses have been detected in some organisms such as the green alga *Halimeda tuna* or its epiphytic hydrozoans. Vagile invertebrates and the fish fauna also show a degree of seasonality, mainly due to recruitment pulses and inactivity in winter. Several suspension feeders also exhibit some physiological seasonality, with decreased activity in summer, probably related to the low food availability and high temperatures that occur during this season.

Some species inhabiting coralligenous assemblages (algae *Mesophyllum alternans*, *Lithophyllum frondosum* and *Halimeda tuna*; sponges *Hemimycale columella*, *Crambe crambe*, *Chondrosia reniformis*, *Dysidea avara* and *Oscarella lobularis*; hydrozoans *Orthopyxis crenata*, *Halecium petrosum* and *H. pusillum*; anthozoans *Paramuricea clavata*, *Eunicella cavolini*, *E. singularis*, *Corallium rubrum*, *Alcyonium acule*, *Parazoanthus axinellae*; tunicates *Halocynthia papillosa*, *Cystodytes dellechiajei* and *Microcosmus sabatieri*) have been carefully studied in order to determine one or several of the following features: growth rates, population dynamics, age, carbonate production, natural diets, prey capture, reproduction, spawning and recruitment patterns.

Five main causes of disturbance that affect coralligenous assemblages have been distinguished:

1. Large-scale events, involving mass mortalities of suspension feeders, seem to be related to summer high water column stability and high temperatures but their ultimate causes remain unclear. It has been suggested that they are related to the current global warming trend.
2. Waste waters profoundly affect the structure of coralligenous communities by inhibiting coralline algal growth, increasing bioerosion rates, decreasing species richness and densities of the largest individuals of the epifauna, eliminating some taxonomical groups (e.g., most echinoderms, bryozoans and crustaceans), and increasing the abundance of highly tolerant species.
3. Fishing is another cause of coralligenous degradation. Trawling is especially destructive, for not only does it physically destroy the coralligenous structure but it also increases turbidity and sedimentation rates, which negatively affects algal growth and suspension feeding. Traditional, as well as recreational, fishing mainly affect target species, although most of them rapidly recover after fishing prohibition or after implementation of scientifically guided fisheries management. However, this is not the case for the long-lived and slow-growing red coral, whose full recovery from harvesting has been estimated to take several decades or even centuries.
4. Diver activity is another cause of recent degradation of coralligenous assemblages, although this kind of disturbance only affects, at the moment, very few areas situated at the most popular sites for recreational diving.

5. Finally, invasive alien species are another cause of concern because their numbers are increasing throughout the Mediterranean. Especially dangerous for the coralligenous communities is the red turf alga *Womersleyella setacea*, which forms a dense carpet over encrusting corallines, thus inhibiting photosynthesis and growth of the main coralligenous builders.

**Actions**

**Gaps in scientific knowledge**

In terms of the current state of scientific knowledge of the coralligenous habitat it is easy to detect several gaps that make it rather difficult to make recommendations for protecting coralligenous assemblages:

1. There is a complete lack of knowledge of the distribution of coralligenous substrata in the Mediterranean, with the exception of some extremely limited areas situated mainly in marine parks or reserves. As a minimum, approximate cartography and quantification of these bottoms is required.

2. It is highly recommended that a list of all the organisms that have been found living in coralligenous communities be drawn up, in order to have a precise idea of the amount of biodiversity contained in this environment.

3. Almost nothing is known about the coralligenous concretions from the eastern Mediterranean. Special efforts must be made to investigate the description and functioning of coralligenous communities in this area.

4. Further studies dealing with the processes involved in the buildup and erosion of coralligenous assemblages must be conducted because almost all the existing information comes from one or two localities situated in the northwestern Mediterranean.

5. An understanding of the functioning of the dominant and keystone species is essential in order to implement an adequate management strategy for the coralligenous habitat.

6. The effect of disturbances in coralligenous assemblages is poorly understood, and there are no data at all on the capacity of this environment to recover (with the exception of fish stocks after fishing prohibition). The following issues would appear to be particularly important:
   a. Indirect impact of trawling
   b. Impact of waste-water dumping
   c. Effects of alien species invasion
   d. Causes of recent large-scale mortality events

**Recommendations for protecting coralligenous communities**

In the light of current knowledge, there are a number of recommendations that can be made in order to conserve (or even improve) coralligenous environments. Most of these recommendations concern not only the coralligenous habitat but most of the coastal benthic habitats because waste-water dumping, trawling and overfishing, and invasion by alien species are problems that affect the whole of the coastal area. Measures to reduce these impacts may improve the overall quality
of the marine coastal environment. Specific measures aimed at protecting the coralligenous environment might include the following:

1. Waste-water dumping should be banned over coralligenous bottoms, and in their vicinity.
2. Trawling must be completely prohibited in areas with coralligenous outcrops and their vicinity, the aim being to avoid not only the physical damage caused by trawling over coralligenous assemblages but also the indirect effects due to increased turbidity and silting.
3. Any other anthropogenic activity involving an increase in water turbidity and/or sediment removal (e.g., coastline modification, beach regeneration) should be avoided in the vicinity of coralligenous outcrops.
4. Correct management of traditional and recreational fisheries must be implemented in order to prevent stock depletion of target fish and crustaceans.
5. The impact of diving must be compatible with the normal functioning and conservation of the coralligenous environment.
6. The enactment of suitable legislation concerning the introduction of alien species is urgently needed.

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MEDITERRANEAN CORALLIGENOUS ASSEMBLAGES


**Colour Figure 7 (Ballesteros)** Types and habitats in coralligenous outcrops. (A) small coralligenous accretion apparently developed from the coalescence of rhodoliths (Tossa de Mar, NE Spain, 40 m depth); (B) coralligenous bank grown upon a rocky outcrop (Tossa de Mar, NE Spain, 25 m depth); (C) community dominated by suspension feeders in a coralligenous cavity (Cabrera, Balearic Islands, 52 m depth); (D) coralligenous rim on a vertical cliff (Gargalo, Corsica, 48 m depth). (Photos by the author.)

**Colour Figure 8 (Ballesteros)** Main red algal building species in coralligenous frameworks. (A) *Mesophyllum alternans*; (B) *Lithophyllum frondosum*; (C) *Lithophyllum cabiochae*; (D) *Neogoniolithon mamillosum*; (E) *Peyssonnelia rosa-marina*. (Photos by the author.)
Colour Figure 9 (Ballesteros) Some animal building species in coralligenous frameworks. (A) *Miniacina miniacea*; (B) *Pentapora fascialis*; (C) *Myriapora truncata*; (D) *Serpula vermicularis*; (E) *Leptopsammia pruvoti*. (Photos by the author.)

Colour Figure 10 (Ballesteros) Bioeroders in coralligenous frameworks. (A) *Cliona viridis*; (B) *Sphaerechinus granularis*; (C) *Echinus melo*; (D) browsing marks of *Sphaerechinus granularis* over *Lithophyllum frondosum*. (Photos by the author.)
Colour Figure 11 (Ballesteros) Diagrammatic section of a coralligenous bank, showing the high small-scale environmental heterogeneity and the different microhabitats. (Drawing by J. Corbera.)

Colour Figure 13 (Ballesteros) Different assemblages of algal-dominated coralligenous banks and rims; (A) with *Halimeda tuna* and *Mesophyllum alternans* (Tossa de Mar, NE Spain, 28 m depth); (B) with *Lithophyllum frondosum* (Tossa de Mar, NE Spain, 40 m depth); (C) with *Peyssonnelia rosa marina*, *Mesophyllum alternans*, *Palmophyllum crassum* and *Peyssonnelia squamaria* (Scandola, Corsica, 50 m depth); (D) detail of C. (Photos by the author.)
Colour Figure 14 (Ballesteros) (A) Drawing of a deep-water, animal-dominated, coralligenous assemblage in the Medes Islands (NE Spain). (B) Key to major species, left from top to bottom: Paramuricea clavata⁶, (and on it Haliotis antillarum⁷), Pteria hirundo⁸, Agkistrodon septifer⁹, Cliona viridis⁵, Alcyonium acaule¹¹, Acanthella acuta¹³, Lithophyllum frondosum⁴, Agelas oroides⁵, Palinurus elephas²⁴, Parazoanthus axinella¹⁹, Spirastrella cunctatrix⁶, Chondrosia reniformis⁸, Petrosea ficiformis⁴ (and on it Smittina cervicornis²² and Discodoris atromaculata²³), Serpula vermicularis²¹, Caryophyllia inornata²⁰, Halocynthia papillosa²⁸, Clathrina coriacea³, Corallium rubrum¹⁹ and Chromis chromis.²² Right, from top to bottom (excluding the above-mentioned species): Anthias anthias³¹, Eunicella singularis¹², Diplodus sargus²³, Codium bursa³, Epinephelus marginatus³⁵, Phyllangia mouchezi³⁵, Galathea strigosa³⁵, Synthecium evansi³⁵, Dystidea avara³⁵. (Drawing by M. Zabala & J. Corbera.)
Different assemblages of animal-dominated coralligenous banks and rims; (A) with gorgonians *Paramuricea clavata* and *Eunicella cavolinii* but also green algae *Halimeda tuna* and *Flabellia petiolata* (Gargalo, Corsica, 45 m depth); (B) with *Paramuricea clavata* and encrusting sponges in deep waters (Cabrera, Balearic Islands, 65 m depth); (C) with sponges, bryozoans and anthozoans (Cabrera, Balearic Islands, 50 m depth); (D) overhangs with *Corallium rubrum* (Palazzu, Corsica, 35 m depth). (Photos by the author.)

Spatial interactions are crucial in the buildup of coralligenous assemblages. (A) *Mesophyllum alternans* overgrows *Lithophyllum cabiochae* which, in its turn, is epiphytised by the small green alga *Halicystis parvula* (above) and a tunicate (below); (B) *Lithophyllum frondosum* overgrows sponge *Ircinia oros*. Strong prey selection is present in the coralligenous community. (C) Opisthobranch *Discodoris atromaculata* feeds almost exclusively on sponge *Petrosia ficiformis*; (D) Opisthobranch *Flabellina affinis* feeds on hydrozoans of the genus *Eudendrium*. (Photos by the author.)
Colour Figure 17 (Ballesteros) (A) Space competition can also be mediated by trophic depletion of the surrounding waters, or by allelochemicals. Tunicate *Pseudodistoma cyrnusense* inhibits growth of bryozoan *Hornera frondiculata*; (B) Zoantharian *Parazoanthus axinellae* is usually a selective epibiont of sponge *Axinella damicornis*; (C) Nonselective epibions overgrow the gorgonian *Paramuricea clavata*: the worm *Salmacina dysteri* and the bryozoan *Pentapora fascialis*; (D) The barnacle *Pyrgoma anglicum* living inside the anthozoan *Leptopsammia pravoti* can be considered a case of parasitism. (Photos by the author.)

Colour Figure 18 (Ballesteros) Maps of transition intensity resulting from overlay procedures of images from the same plot (310 cm²) along a depth gradient in a vertical wall at the Medes Islands (NE Spain) during 2 yr of sampling. Patch colour denotes number of changes taking place in each patch (see legend). Coralligenous communities (14 and 20 m depth) display much lower transition rates than shallow water communities, indicating high persistence and low rates of change in the animals and plants thriving in the coralligenous communities. (From Garrabou et al. 2002. With permission from Elsevier.)
Colour Figure 22 (Ballesteros) A red coral colony (age unknown) from a pristine site (Cap Creus, 35 m) collected in 1962 (A), and a 28-year-old colony from an experimental panel (Riou Caramassaigne, 62 m) (B). (Photo and data courtesy of J.G. Harmelin.)

Colour Figure 25 (Ballesteros) Disturbances in coralligenous communities. (A) Mortality affecting the gorgonian Paramuricea clavata (Port-Cros, France, autumn 1999); (B) Dense carpets of alien alga Womersleyella setacea cover coralligenous assemblages; the gorgonian Eunicella singularis is also affected by previous partial mortality that occurred in summer 1999 (Minorca, Balearic Islands, summer 2000); (C) Filamentous alien alga Womersleyella setacea invades coralligenous rims dominated by Mesophyllum alternans in Cabrera (Balearic Islands, autumn 1999); (D) Partial mortality and overgrowth by filamentous algae affecting gorgonian Eunicella singularis (Minorca, Balearic Islands, summer 2000). (Photos by the author.)
**Colour Figure 2 (Collins & Villanueva)** Photographs of cirrate octopods. (A) dorsal view of *Opisthoteuthis massyae* (fresh specimen), (B) ventral view of *Cryptoteuthis brevibracchiata* (fresh specimen), (C) dorso-posterior view of *Cirroctopus glacialis* (fresh specimen), (D) ventral view of *Grimpoteuthis discoveryi* (formalin-preserved specimen), (E) ventral view of *Cirrothauma murrayi* (fresh specimen), (F) oral view of male *Stauroteuthis syrtensis* (formalin-preserved specimen), (G) Juvenile specimen of *Opisthoteuthis calypso*, note the relatively large fins and funnel in comparison with the adult *Opisthoteuthis* in (A). Sources (with permission where required): (A) Collins unpublished; (B) from Collins (2004); (C) Mike Vecchione unpublished; (D) from Collins (2003); (E) from Aldred et al. (1983); (F) from Collins & Henriques (2000); (G) L. Dantart. Scale bars: (A–F) = 100 mm; (G) = 10 mm.

**Colour Figure 20 (Collins & Villanueva)** Photographs of cirrate octopods taken from the manned submersible *NAUTILE* at the mid-Atlantic Ridge, illustrating behaviour. *Grimpoteuthis* sp.: (A) bottom resting and (B, C) crawling. Cirroteuthidae: (D) drifting in umbrella style, (E) during take off and (F) fin-swimming. *Grimpoteuthis* sp.: (G–I) fin-swimming. Cirroteuthidae: (J) umbrella drifting, (K, L) taking off after touching submersible and displaying long cirri, (M, N) swimming by pumping. (O) *Cirrothauma magna* being manoeuvred into a sample box, showing ballooning response in three web sectors. (From Villanueva et al. (1997). With permission.)